

New cancer diagnostic technique debuts

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Cancer cells break down sugars and produce the metabolic acid lactate at a much higher rate than normal cells. This phenomenon provides a telltale sign that cancer is present, via diagnostics such as PET scans, and possibly offers an avenue for novel cancer therapies. Now a team of Chilean researchers at The Centro de Estudios Científicos (CECs), with the collaboration of Carnegie's Wolf Frommer, has devised a molecular sensor that can detect levels of lactate in individual cells in real time.

Prior to this advance, no other <u>measurement method</u> could noninvasively detect lactate in real time at the single-cell level. The work, published in the open access journal *PLOS ONE*, is a boon to understanding how different types of cells go awry when cancer hits.

"Over the last decade, the Frommer lab at Carnegie has pioneered the use of Förster <u>Resonance Energy Transfer</u>, or FRET, sensors to measure the concentration and flow of sugars in individual cells with a simple fluorescent color change. This has started to revolutionize the field of <u>cell metabolism</u>," explained CECs researcher Alejandro San Martín, lead author of the article. "Using the same underlying <u>physical principle</u> and inspired by the sugar sensors, we have now invented a new type of sensor based on a transcriptional factor. A molecule that normally helps bacteria to adapt to its environment has now been tricked into measuring lactate for us."

Lactate shuttles between cells and inside cells as part of the normal <u>metabolic process</u>. But it is also involved in diseases that include inflammation, inadequate <u>oxygen supply</u> to cells, restricted <u>blood supply</u>



to tissues, and neurological degradation, in addition to cancer.

"Standard methods to measure lactate are based on reactions among enzymes, which require a large number of cells in complex cell mixtures," explained Felipe Barros, leader of the project. "This makes it difficult or even impossible to see how different types of cells are acting when cancerous. Our new technique lets us measure the metabolism of individual cells, giving us a new window for understanding how different cancers operate. An important advantage of this technique is that it may be used in high-throughput format, as required for drug development."

This work used a bacterial transcription factor—a protein that binds to specific DNA sequences to control the flow of genetic information from DNA to mRNA—as a means to produce and insert the lactate sensor. They turned the sensor on in three cell types: normal brain cells, tumor brain cells, and human embryonic cells. The sensor was able to quantify very low concentrations of lactate, providing an unprecedented sensitivity and range of detection.

The researchers found that the tumor cells produced lactate 3-5 times faster than the non-tumor cells. "The high rate of lactate production in the cancer cell is the hallmark of cancer metabolism," remarked Frommer. "This result paves the way for understanding the nuances of cancer metabolism in different types of cancer and for developing new techniques for combatting this scourge."

In addition, the biosensors promise to solve an old controversy. While some studies have suggested the glucose provides the fuel for the brain, recent research has provided evidence that lactate feeds energy metabolism in neurons. Oxidation of lactate can be used to produce large amount of ATP—the coenzyme that carries energy in cells. The Barros and Frommer teams are excited about the solving this enigma with the use of their new sensors, together with the previously developed glucose



sensors. Recently, a collaboration between the two labs led to the patenting of the first method capable of measuring the rate of glucose consumption in single cells.

Provided by Carnegie Institution for Science

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