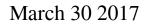


Resilient red blood cells need fuel to adapt their shape to the environment



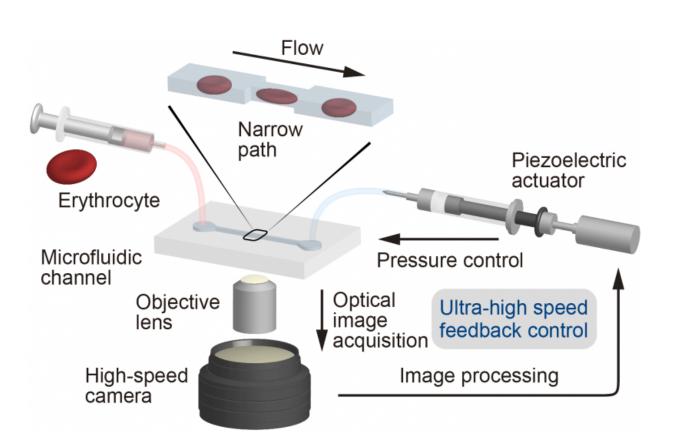


Figure 1: Microchannel connected to a robotic pump. Feedback loop between high-speed vision and actuator enables real-time on-chip manipulation of a red blood cell (RBC). RBC deforms inside the narrow path for a specified time. Credit: Osaka University

An international research team led by Osaka University built a novel



"Catch-Load-Launch" microfluidic device to monitor the resilience of red blood cells after being held in a narrow channel for various periods of time. They found that the time for the red blood cell to spring back into shape was shorter for when starved of adenosine triphosphate or exposed to endotoxins. These findings may help improve treatments for patients with sepsis or malaria.

Red blood <u>cells</u>, also called erythrocytes, need a certain springiness to circulate through the body. Some capillary blood vessels are smaller than the diameter of an erythrocyte, which forces them to be folded as they squeeze through. Researchers have developed a new system for studying the resilience of red blood cells. This finding may lead to improved diagnosis and treatment of blood-related diseases, for example, <u>septic</u> <u>shock</u> and malaria.

To study how erythrocytes spring back into shape after deforming to pass through a constriction, an international team of researchers centered at Osaka University built what is called a "Catch-Load-Launch" microfluidic platform. The findings were recently published in *Scientific Reports*.

The experimental setup included a microchannel, in which a single erythrocyte could be held in place for any desired length of time before being launched into a wider section using a robotic pump, simulating the transition from a capillary into a larger vessel.

"The cell was precisely localized in the microchannel by the combination of pressure control and real-time visual feedback," study coauthor Makoto Kaneko says. "This let us 'catch' an erythrocyte in front of the constriction, 'load' it inside for a desired time, and quickly 'launch' it from the constriction to monitor the shape recovery over time."



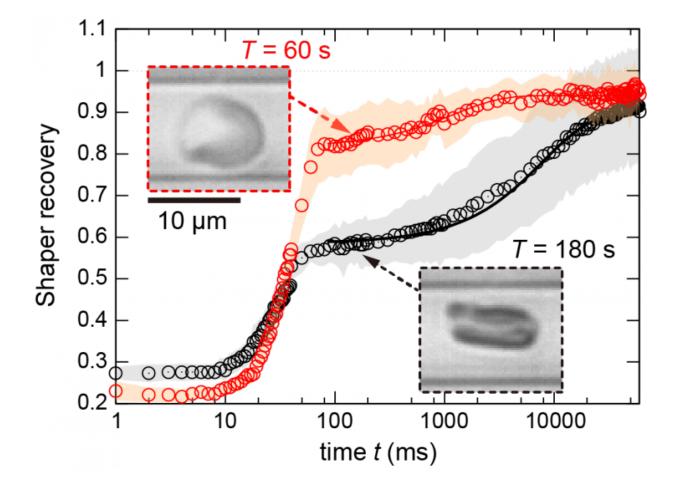


Figure 2: Drastic difference in red blood cell (RBC) deformability in response to different loading times. The shape recovery after 180 s deformation is drastically slower, indicating the internal remodeling of cytoskeletons. Credit: Osaka University

Each erythrocyte has an internal cytoskeleton made of interlinking networks of filaments, including spectrin proteins, that contributes to its rigidity. Dynamic remodeling of this scaffold as the erythrocyte changes shape requires the cell to expend fuel, usually adenosine triphosphate (ATP). The researchers found that as the time the erythrocyte was held in the constricted region was increased—from 5 seconds all the way to 5 minutes—the time it took the cell to recover its normal shape also



increased. For very short constriction times, the cells bounced back within 1/10 of a second. Conversely, it took approximately 10 seconds for cells to recover if they were held in the narrow segment longer than about 3 minutes.

The researchers next studied how the red blood cells' recovery time was affected by a lack of ATP. Contrary to intuition, cells deprived of this fuel, ATP, took less time to recover.

Finally, the "Catch-Load-Launch" system was used to study the lifethreatening condition known as septic shock. This can occur when bacteria invade the bloodstream and release endotoxins, which are known to affect cytoskeleton proteins. Patients in septic shock may suffer from reduced circulation inside the narrow blood vessels as the erythrocytes become too stiff. The same problem can be caused by Plasmodium falciparum, the parasite responsible for malaria.

The researchers exposed red blood cells to endotoxin from the bacteria Salmonella minnesota, and found the erythrocytes became stiffer and less resilient, like those starved of ATP.

"There is a great deal of evidence that relates certain diseases, including sepsis and malaria, to a decrease in the deformability of red blood cells," lead author Hiroaki Ito says. "Such a stiffening can lead to a disturbance in microcirculation, and our 'Catch-Load-Launch' platform has the potential to be applied to the mechanical diagnosis of these diseased <u>blood</u> cells."

More information: Hiroaki Ito et al. Mechanical diagnosis of human erythrocytes by ultra-high speed manipulation unraveled critical time window for global cytoskeletal remodeling, *Scientific Reports* (2017). DOI: 10.1038/srep43134



Provided by Osaka University

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