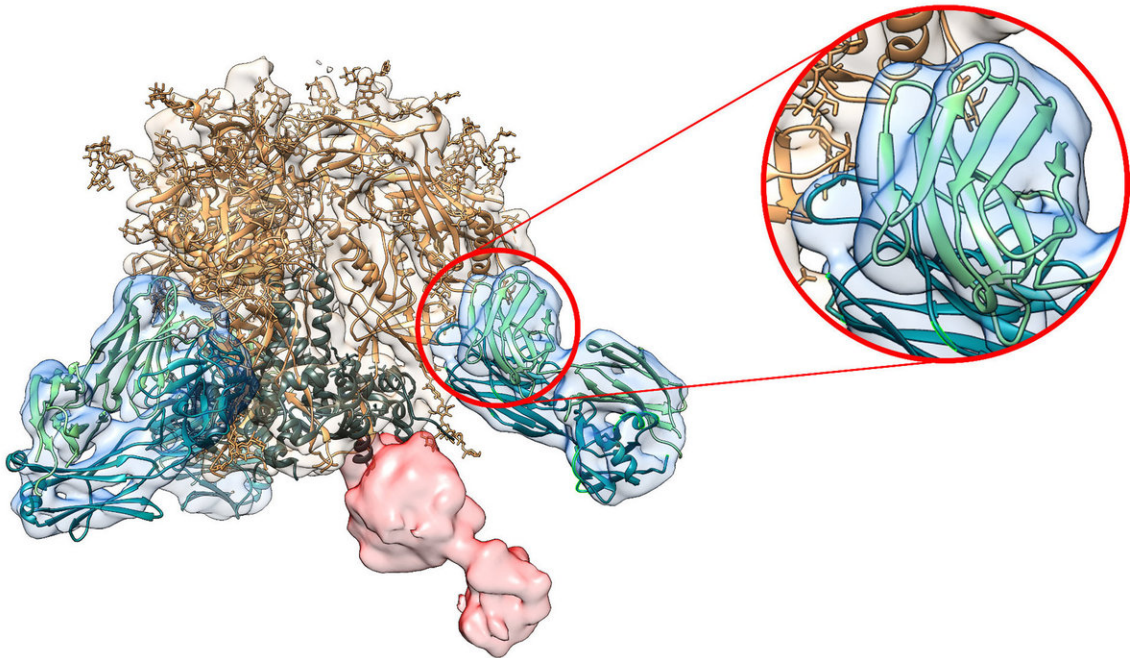


New antibody analysis accelerates rational vaccine design

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With the new method, the researchers were able to image polyclonal antibody/HIV envelope complexes at 4.7Å resolution. At this resolution, the researchers discovered that in rabbits, antibodies specific to a vulnerable sites on HIV's outer glycan layer (blue shapes) are structurally highly convergent and closely resemble a previously isolated monoclonal antibody 10A (green and teal ribbons). Credit: Hangartner, Ward labs, Scripps Research

A team at Scripps Research has come up with a faster way to analyze the

outcome of experimental vaccines against HIV and other pathogens. Their new system lets scientists quickly assess the full spectrum of antibodies produced in an individual in response to a pathogen or vaccine and determine if these antibodies are likely to be effective against the pathogen.

"We can now watch antibody responses evolve almost in real time," says Lars Hangartner, Ph.D., a Scripps Research associate professor and co-senior author of the study, which published today in the journal *Immunity*.

"This can be applied to any pathogen," adds Andrew Ward, Ph.D., professor at Scripps Research and study co-senior author. Ward is also affiliated with the International AIDS Vaccine Initiative (IAVI) Neutralizing Antibody Center and Center for HIV/AIDS Vaccine Immunology and Immunogen Discovery (CHAVI-ID) at Scripps Research.

When a pathogen attacks, your immune system amasses an army of [antibodies](#) to fight back. Some of these antibodies will fail, but some may get close to beating a pathogen's defenses. Over time, the best antibodies will evolve to target a pathogen's vulnerable sites. If all goes well, antibodies will home in on those epitopes, neutralize the invader and provide lifelong immunity.

Vaccines work by tricking the body into producing neutralizing antibodies against a future invader. Testing vaccines tends to be a lengthy process, with an initial immunization and then a series of booster shots over weeks or months. The time it takes to analyze the trial data means scientists don't usually get to see if a vaccine is working until the experiment is over, making it especially hard to tweak treatment strategies in the face of emerging diseases.

With the new study, scientists finally have a way to get a nearly real-time picture of antibody evolution. The new technique builds on Scripps Research-led breakthroughs in immunology and an imaging technique called electron microscopy, which reveals the structures of antibodies bound to their target [pathogens](#), such as HIV.

The researchers used rabbit blood samples collected at different stages of an HIV vaccine trial. The researchers purified the samples, breaking down the molecules until they could extract antibody fragments. They then mixed these antibody fragments with their targets-viral proteins-and imaged them in the electron microscope, revealing how the immune system was developing its attack on the pathogen.

"After fine-tuning the biochemistry behind the formation of immune complexes of HIV Env trimers with antibody fragments, we were able to establish a very solid protocol allowing for semi-quantitative electron microscopy analysis," says study co-first author Matteo Bianchi, Ph.D., a former Scripps researcher now with the Institute of Medical Virology, University of Zurich.

This new method is based on images taken by a relatively low-tech method called negative stain imaging, which helped them spot promising, or more often than not, distracting, antibodies. At last, the researchers could quickly find out if a vaccine was pushing the immune system along the right path. And in cases where a vaccine wasn't working, the method could provide researchers with information as to how the vaccine could be improved.

"You can change the direction of a study depending on the results from this method," says study co-first author Hannah Turner, a research assistant in the Ward lab. "Never before have we been able to take a snapshot of our antibodies like this-giving us a more complete picture of what is happening and how we can influence the vaccination process."

While the images were not high-resolution, the researchers were excited to report that this level of analysis-which only takes days-could become a routine test around the world. "Most institutes have at least the technology and resources to do this level of analysis," says Ward.

For an even closer look at the antibodies, Ward used higher-tech cryo-electron microscopy to get high-resolution, 3-D images of the antibodies with their viral targets. These detailed snapshots revealed additional details that may help scientists improve future experimental HIV vaccines.

The researchers say their faster method could open the door to personalized design for vaccines. For example, this study included a "high-responder" rabbit that produced many effective antibodies, and a "low-responder" rabbit with a weak army of antibodies. Going forward, researchers could use this new method in human [vaccine](#) or therapeutic trials to quickly separate high responders from low responders early on, saving crucial time for patients and their doctors as they search for effective therapies and vaccines that provide herd immunity.

"This is going to be part of every immunization project I do from now on," Ward says.

"This method has the potential to change the pace at which we can develop vaccines," adds Hangartner.

More information: Matteo Bianchi et al, Electron-Microscopy-Based Epitope Mapping Defines Specificities of Polyclonal Antibodies Elicited during HIV-1 BG505 Envelope Trimer Immunization, *Immunity* (2018). [DOI: 10.1016/j.immuni.2018.07.009](https://doi.org/10.1016/j.immuni.2018.07.009)

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