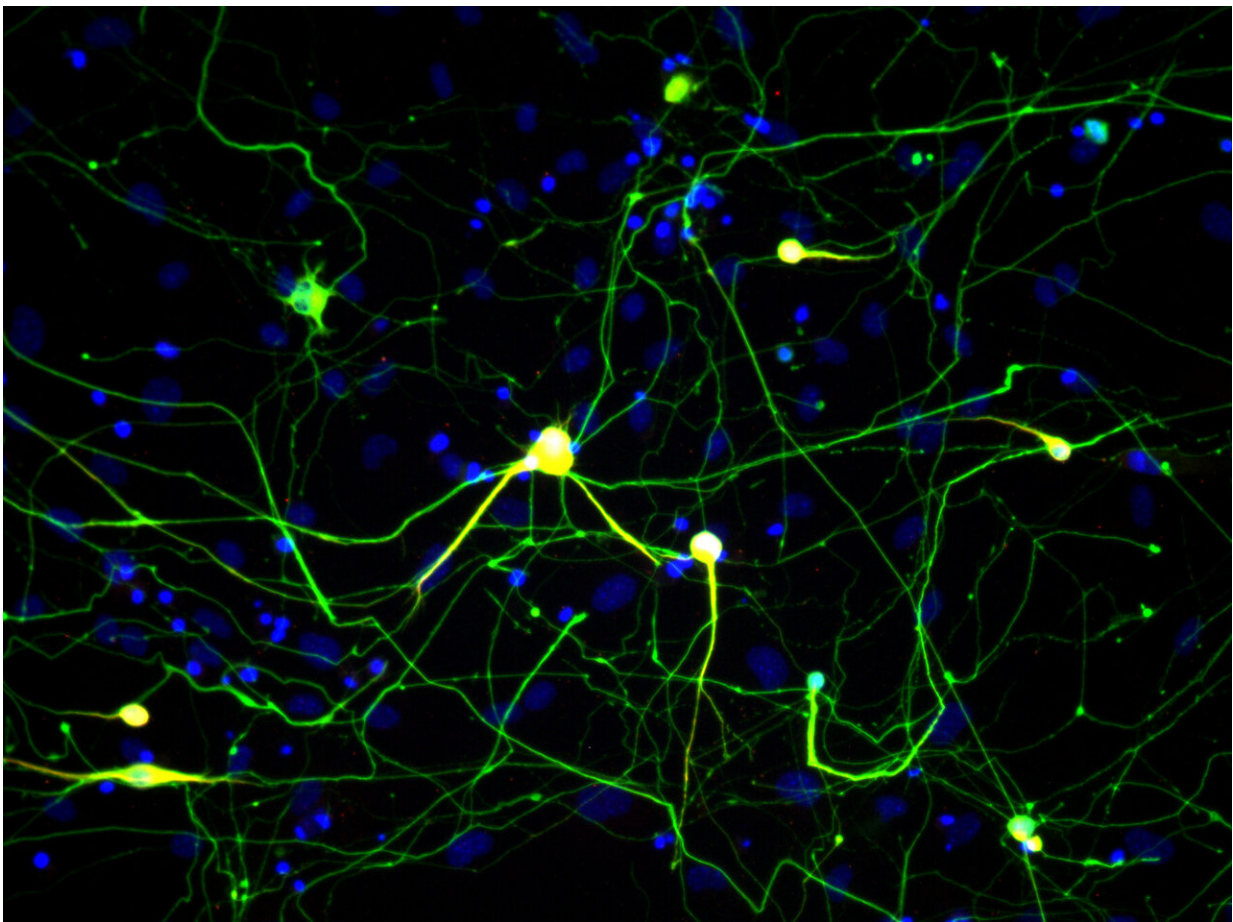


Research confirms nerve cells made from skin cells are a valid lab model for studying disease

January 15 2019



Induced neuronal cells. Credit: Wernig lab/Stanford University

The incidence of some neurological diseases—especially those related to aging, such as Alzheimer's and Parkinson's diseases—is increasing. To better understand these conditions and evaluate potential new treatments, researchers need accurate models that they can study in the lab.

Researchers from the Salk Institute, along with collaborators at Stanford University and Baylor College of Medicine, have shown that cells from mice that have been induced to grow into [nerve cells](#) using a previously published method have molecular signatures matching [neurons](#) that developed naturally in the brain.

The study, published in *eLife* on January 15, 2019, opens the door for better ways to model an individual patient's disease. This technique would enable researchers to study how neurological conditions develop, as well as to test new therapies. The [new technology](#) also could help to advance research into gene therapies that are derived from a patient's own cells.

"This research is charting the path for the most optimal way of creating neurons in the lab," says Salk Professor Joseph Ecker, one of the study's two senior authors. "By taking these cells and reprogramming them into neurons, you can potentially learn new things about how these diseases function on a cellular level, especially diseases driven by genetic changes."

The cells used in the study, called fibroblasts, make up most of the [connective tissue](#) in animals and play an important role in wound healing. Researchers have been studying how to transform fibroblasts into neuron cells in laboratory dishes, but until now they didn't know whether these newly created neurons accurately corresponded to neurons that had grown naturally in the brain.

The technique for inducing the fibroblasts to grow into neurons with the

matching epigenome was developed by Stanford's Marius Wernig, the paper's co-senior author. With this method, making induced [neuronal cells](#) does not involve pluripotent intermediates. Instead, the cells are directly converted from fibroblasts to neurons.

"An important question in cellular engineering is how to know the quality of your product," says co-first author Chongyuan Luo, a postdoctoral fellow in Ecker's lab. "If we're making neurons from fibroblasts, we want to know how they compare with neurons in the brain. We are particularly interested in looking at these cells at the level of the epigenome."

The epigenome is made up of chemicals that attach to DNA and regulate when genes get turned on and translated into proteins. Differences between the epigenomes of induced and naturally grown neurons could result in different features of induced neurons that might make them less accurate models of neuronal behavior.

Using a technique developed in the Ecker lab called MethylC-seq, the researchers looked at every place in the genome where chemical groups called methyl groups are attached. They confirmed that these induced neurons have epigenomes that match neurons in the brain.

"This research was done in mouse cells, but we plan to use the same technology to study induced neurons made with human [cells](#)," explains Ecker, who is director of Salk's Genomic Analysis Laboratory and a Howard Hughes Medical Institute investigator. Ecker plans to also collaborate with colleagues to apply the technology to look at [human cells](#) to better understand age-related cognitive decline.

More information: Chongyuan Luo et al, Global DNA methylation remodeling during direct reprogramming of fibroblasts to neurons, *eLife* (2019). [DOI: 10.7554/eLife.40197](https://doi.org/10.7554/eLife.40197)

Provided by Salk Institute

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