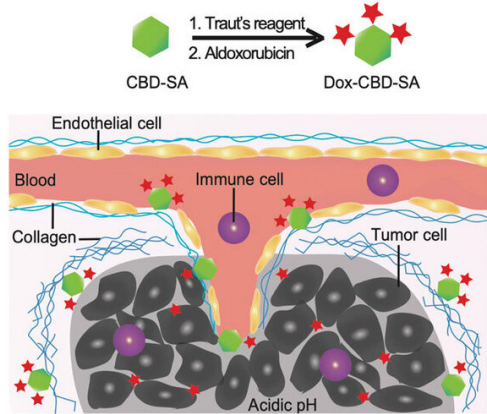


# **Engineering collagen-binding serum albumin (CBD-SA) as a drug conjugate carrier for cancer therapy**

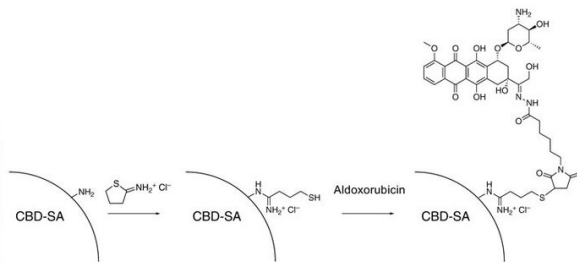
August 23 2019, by Thamarasee Jeewandara

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**A**



**B**



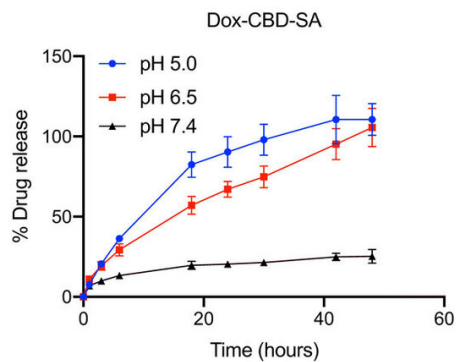
**C**

$K_d$ (nM)	Collagen I	Collagen III
CBD-SA	1.8	40.6
SA	N.D.	N.D.

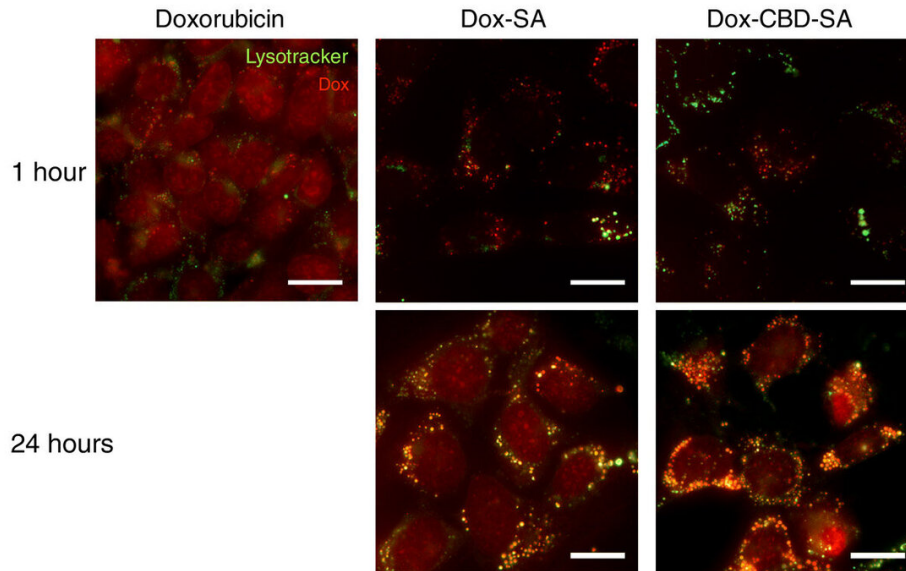
**D**

	Dox:protein ratio
Dox-SA	$3.1 \pm 0.1$
Dox-CBD-SA	$3.4 \pm 0.1$

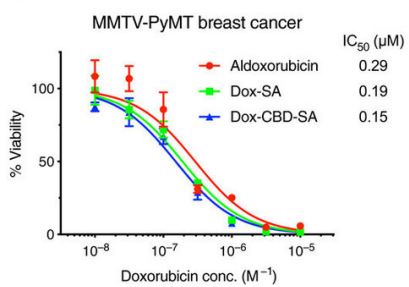
**E**



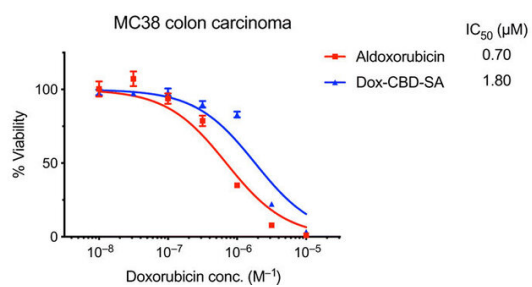
**F**



**G**



**H**



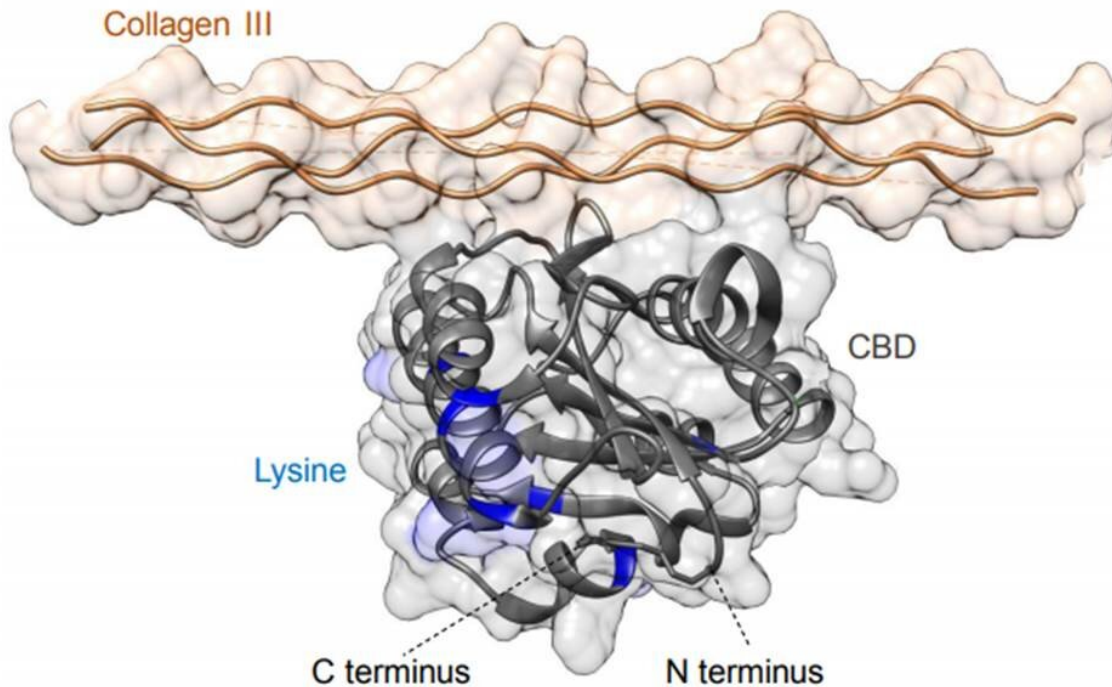
Synthesis and characterization of Dox-CBD-SA. (A) Schematic of CBD-SA mediated drug delivery. (B) Synthesis scheme of Dox-CBD-SA. (C) Affinities [dissociation constant ( $K_d$ ) values are shown] of CBD-SA and SA against collagen type I and collagen type III were measured by enzyme-linked immunosorbent assay (ELISA). N.D., not determined due to low signal. (D) Dox conjugation ratio per protein is presented. Values were calculated on the basis of the results of bicinchoninic acid assay protein quantification assay (proteins) and absorbance at 495 nm (Dox) (mean  $\pm$  SD of three experimental replicates). (E) Dox release kinetics from Dox-CBD-SA under three different pH conditions was evaluated by fluorescence (excitation at 495 nm, emission at 590 nm) ( $n = 3$ , mean  $\pm$  SD; two experimental replicates). (F) MMTV-PyMT cells were seeded and incubated overnight. Dox, Dox-SA, or Dox-CBD-SA was added (red). Cells were also stained with LysoTracker (green). Scale bars, 20  $\mu$ m. Representative pictures are presented. Two experimental replicates. (G and H) Cytotoxicity of Dox variants against MMTV-PyMT cells or MC38 cells in vitro ( $n = 6$ , mean  $\pm$  SEM). Credit: *Science Advances*, doi: 10.1126/sciadv.aaw6081

Medical researchers often use serum albumin (SA) as a drug carrier to deliver cytotoxic agents to tumors during biomedical drug delivery via passive targeting approaches. To improve the targeting capacity of SA's a team of scientists recently developed an approach to retain SA-drug conjugates in tumors by combining passive and active targeting mechanisms. In the new study, Koichi Sasaki and colleagues in the Pritzker School of Molecular Engineering in the University of Chicago, U.S., recombinantly fused SA with a collagen-binding domain (CBD) of the von Willebrand factor protein. The approach allowed binding within the tumor stroma after drug release, due to tumor-vascular permeability. The work is now published on *Science Advances*.

The research team then conjugated doxorubicin (Dox; an anticancer

[drug](#)) to the CBD-SA conjugate using a pH-sensitive linker. The Dox-CBD-SA treatment significantly suppressed [tumor](#) growth compared to Dox-SA and [aldoxorubicin](#) treatment in a mouse model of breast cancer. Notably, the new compound—Dox-CBD-SA, efficiently stimulated host anti-tumor immunity to completely eradicate established tumors in an animal model of [MC38 colon carcinoma](#) when the research team combined the compound with the [anti-PD1 checkpoint inhibitor](#). The compound DOX-CBD-SA decreased adverse side-effects compared to aldoxorubicin, confirming the bioengineered CBD-SA as a versatile, clinically relevant drug conjugate carrier protein with potential to treat solid tumors.

Serum albumin is the [most abundant protein in blood](#) and a number of small molecule compounds can be fused, conjugated with, or co-formulated with SA for improved drug delivery to disease lesions. The exceptionally long plasma half-life and [hydrophilicity](#) (water loving nature) of SA contributed to [improve the pharmacokinetics](#), safety and [efficacy](#) of the [new drug compounds](#). Notably, SA can passively target tumors via pathological permeability of the tumor vasculature, which is advantageous for cancer therapy. Molecular engineering approaches have gained prominence in the field of cancer research to combine passive and active targeting [during tumor drug delivery](#).



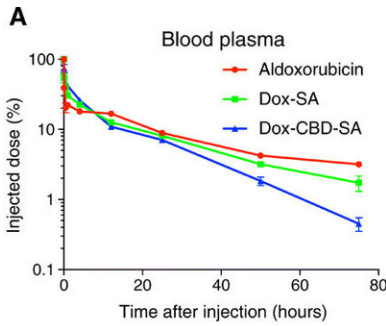
The binding interface between collagen type III and A3 domain of VWF. Crystal structure of the A3 domain of von Willebrand factor (CBD) in complex with type III collagen (PDB 4DMU). The Image was processed using UCSF chimera. Lysines are indicated as blue color. Credit: Science Advances, doi: 10.1126/sciadv.aaw6081

Sasaki and team had [previously shown](#) the targeted delivery of [checkpoint inhibitor](#) (CPI) antibodies and the [cytokine interleukin-2](#) (IL-2) using a [collagen binding domain](#) (CBD) known as the [A3 domain](#) of von Willebrand factor (VWF). The A3 domain of VWF has [the highest affinity to collagen](#) types I and III proteins. This is since collagen is [abnormally expressed](#) and exposed to the bloodstream due to hyperpermeability of the tumor vasculature, forming a promising target for cancer drug delivery. The bioengineered forms in the preceding

study showed significantly stronger anti-tumor effects compared to their unmodified forms [in mouse models of cancer](#). Researchers had also suppressed treatment-related adverse events by fusing CBD to the drug during the tumor targeting strategy. In the present work, Sasaki et al. hypothesized that CBD would be similarly compatible with SA-based drug delivery carriers to engineer an active serum albumin (SA) with tumor targeting potential.

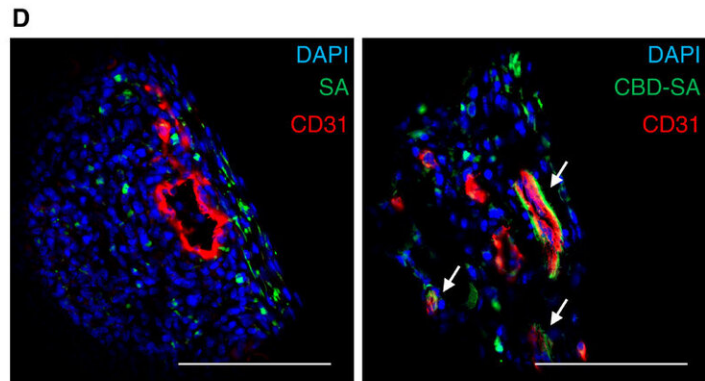
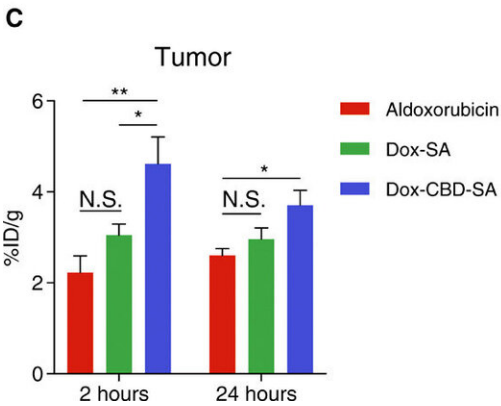
The research team focused on Doxorubicin (Dox), a small-molecule anticancer drug approved by the US Food and Drug Administration (FDA) to treat a broad spectrum of cancer, where the molecule can internalize in proliferating cells through passive transmembrane diffusion to interfere with DNA functions and cause cell death. However, its antitumor efficacy is not notable due to [acquired drug resistance](#), bone marrow suppression and [excessive inflammation](#) among patient populations, leading to a poor therapeutic index. To improve Dox efficacy, researchers often combine other chemotherapeutic agents, such as synergized treatment with CPI (checkpoint inhibitors) or Dox conjugated with SA for its release in the low pH (reportedly pH 6.5) tumor microenvironment [in a mouse model](#).





**B**

	$t_{1/2, \alpha}$ (hours)	$t_{1/2, \beta}$ (hours)
Aldoxorubicin	$4.0 \times 10^{-2} \pm 2.8 \times 10^{-3}$	$22.6 \pm 3.3$
Dox-SA	$5.0 \times 10^{-2} \pm 7.8 \times 10^{-3}$	$8.4 \pm 1.7$
Dox-CBD-SA	$9.0 \times 10^{-2} \pm 2.4 \times 10^{-2}$	$6.4 \pm 1.3$



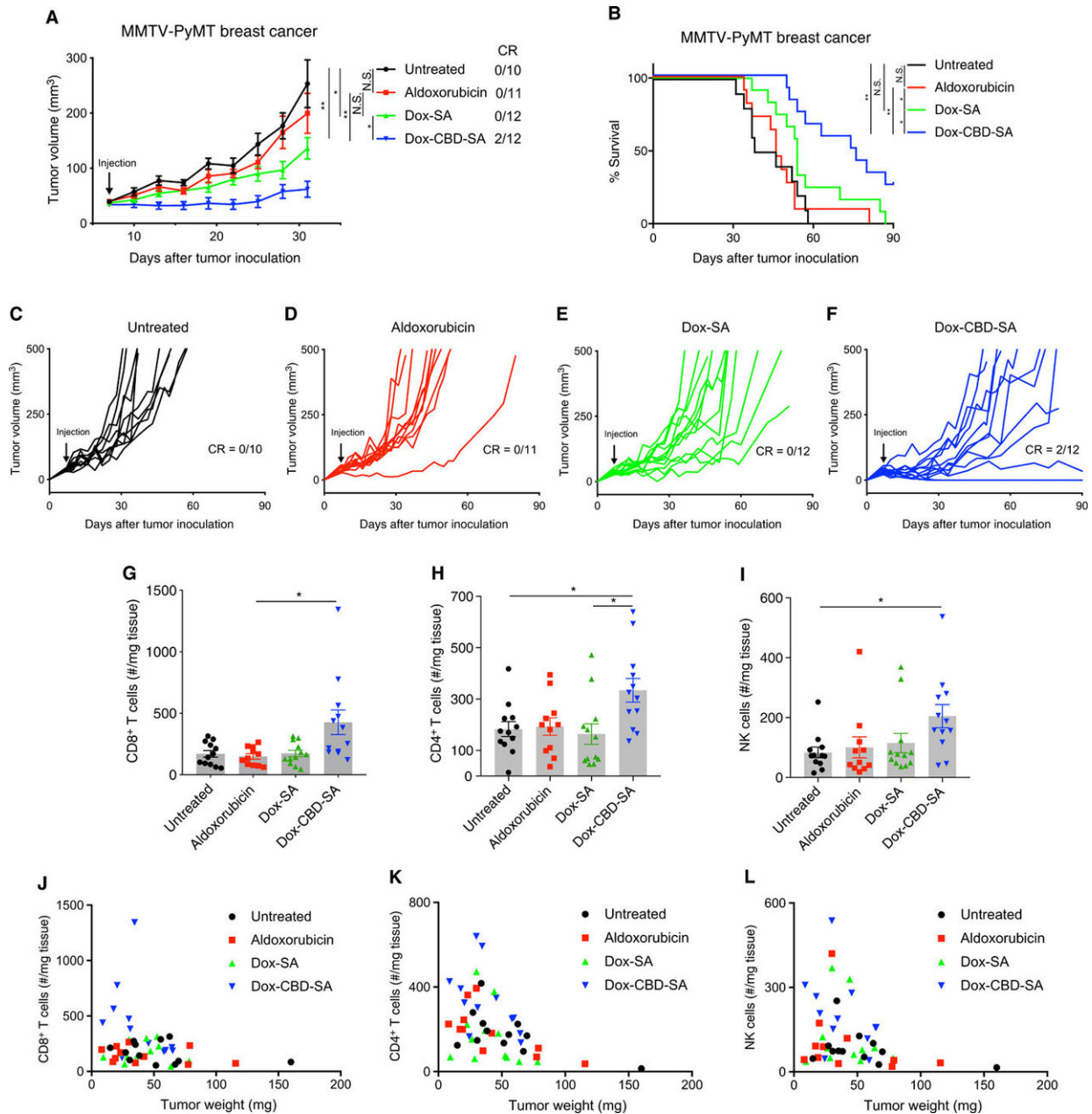
Dox-CBD-SA shows comparable plasma pharmacokinetics with Dox-SA and higher tumor accumulation than aldoxorubicin and Dox-SA. (A) Aldoxorubicin, Dox-SA, or Dox-CBD-SA (5 mg/kg on a Dox basis) was administered to tumor-free FVB mice via tail vein injection. Blood plasma was collected at the indicated time points. Plasma concentration of Dox was measured by fluorescence (mean  $\pm$  SEM;  $n = 4$  for aldoxorubicin,  $n = 5$  for Dox-SA and Dox-CBD-SA). (B) Plasma half-lives of Dox were calculated using two-phase exponential decay:  $MFI(t) = Ae^{-\alpha t} + Be^{-\beta t}$ .  $t_{1/2, \alpha}$ , fast clearance half-life;  $t_{1/2, \beta}$ , slow clearance half-life (mean  $\pm$  SEM;  $n = 4$  for aldoxorubicin,  $n = 5$  for Dox-SA and Dox-CBD-SA). (C) MMTV-PyMT tumor-bearing mice were treated with aldoxorubicin, Dox-SA, or Dox-CBD-SA (4.16 mg/kg on a Dox basis). At the indicated time points, tumors were harvested, and the amount of Dox within the tumors was quantified (mean  $\pm$  SEM;  $n = 5$  for 2 hours,  $n = 7$  for 24 hours per group). (D) DyLight 488-labeled SA (100  $\mu$ g) or equimolar amounts of DyLight 488-labeled CBD-SA were injected intravenously to MMTV-PyMT tumor-bearing mice. One hour after injection, tumors were harvested and fluorescence was analyzed by confocal microscopy. Tissues were also stained with 4',6-diamidino-2-phenylindole (DAPI) and anti-CD31 antibody. Scale bars, 100  $\mu$ m. Representative images of three tumors each. Two experimental

replicates. Statistical analyses were done using analysis of variance (ANOVA) with Tukey's test. \*P

In the present work, Sasaki et al. designed a recombinant mouse SA by fusing the collagen binding domain of the VWF A3 domain (CBD-SA). Then they conjugated doxorubicin (derivative of Dox) to CBD-SA using a pH-dependent [cleavable hydrazone link](#), prior to experimental injection as the "Dox-CBD-SA" therapeutic agent. The research team tested the engineered CBD-SA as a tumor-targeting drug carrier for improved antitumor efficacy with Dox, in the tumor microenvironment of a translational animal model.

The scientists first synthesized the new drug conjugates to target the tumor microenvironment and investigated the binding potential of CBD-SA to the recombinant collagen protein in vitro to show strong binding affinities to collagen types I and III. They covalently conjugated doxorubicin to CBD-SA and conducted [SDS-polyacrylamide gel electrophoresis \(SDS-PAGE\)](#) to observe the monomeric structure of purified DOX-SA and DOX-CBD-SA molecules. They examined the release kinetics of Dox from conjugates under varying pH conditions to show maximum release at pH 5.0 and 6.5, consistent with previous reports on [small chemical release kinetics](#) in tumor microenvironments.



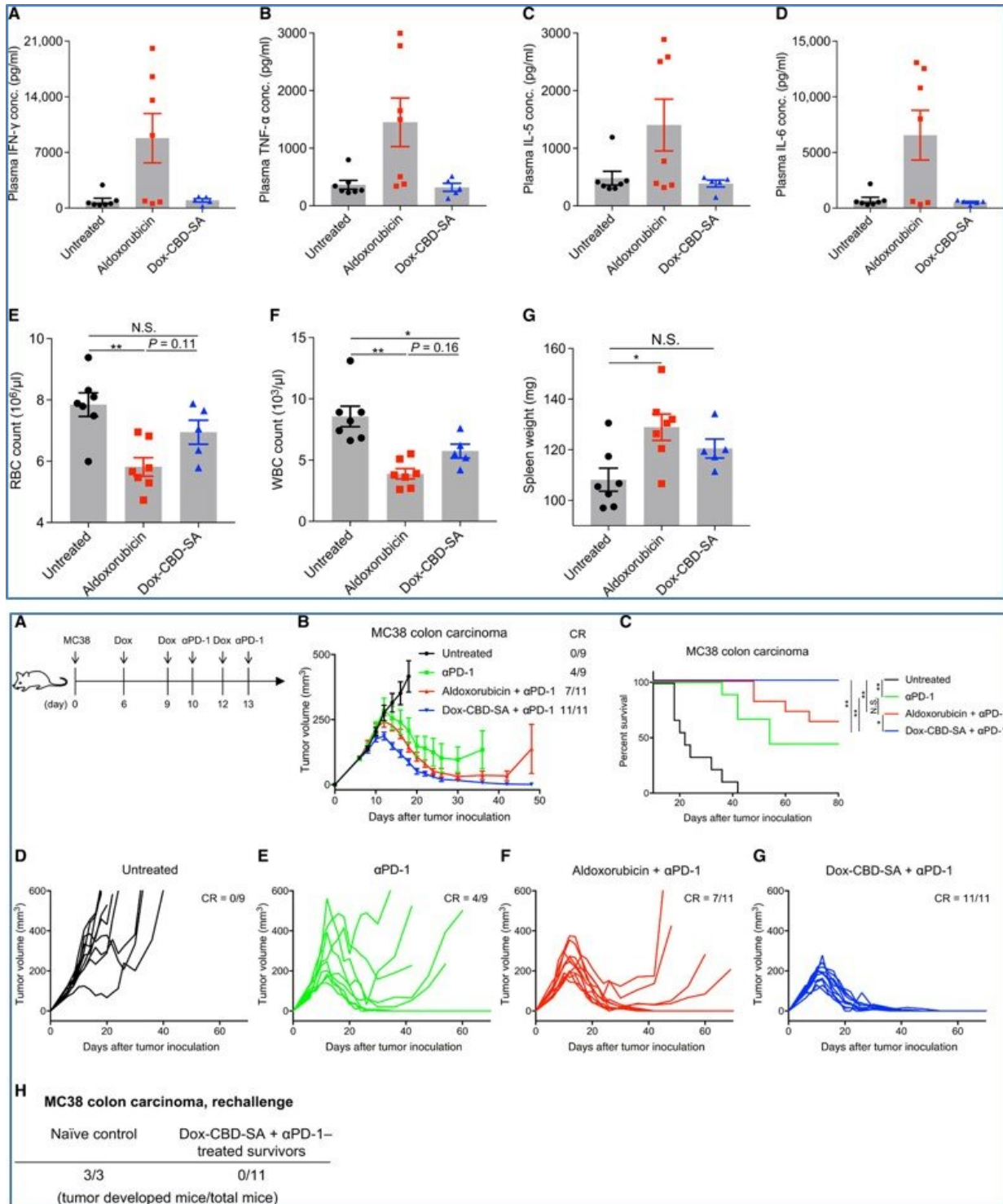


Dox-CBD-SA shows enhanced antitumor efficacy and infiltration of lymphocytes into tumor in the MMTV-PyMT breast cancer model. (A) MMTV-PyMT cells ( $5 \times 10^5$ ) were inoculated into FVB mice on day 0. Aldoxorubicin, Dox-SA, or Dox-CBD-SA (5 mg/kg on a Dox basis) was injected intravenously on day 7. Graphs depict tumor volume until the first mouse died (mean  $\pm$  SEM). (B) Survival rate. (C to F) Individual tumor growth curves. CR indicates complete response frequency. Three experimental replicates. (G to L) MMTV-PyMT cells ( $5 \times 10^5$ ) were inoculated on day 0. Aldoxorubicin, Dox-SA, or Dox-CBD-SA (5 mg/kg on a Dox basis) was injected intravenously on day 7. Lymphocytes within tumors were extracted on day 14, followed by flow cytometric

analysis. (G to I) Graphs depict the number of (G) CD45+CD8+CD3+ T cells, (H) CD45+CD4+CD3+ T cells, and (I) CD45+NK1.1+CD3- NK cells per tumor weight (in milligrams). Bars represent mean  $\pm$  SEM. (J to L) Graph shows [CD45+CD8+CD3+ T cells per tumor weight (mg)] (J), [CD45+CD4+CD3+ T cells per tumor weight (mg)] (K), or [CD45+NK1.1+CD3- NK cells per tumor weight (mg)] (L) versus [tumor weight]. Two experimental replicates. Statistical analyses were done using (A, H, and I) ANOVA with Tukey's test or (G) Kruskal-Wallis test followed by Dunn's test or (B) log-rank (Mantel-Cox) test. \*P

Using cell culture studies the team detected the presence of Dox in the cytoplasm of [MMTV-PyMT](#) cells (mouse mammary tumor virus-polyomavirus middle T antigen) after 1-hour of incubation. After 24-hours of incubation with Dox-conjugates they noted the uptake of the liberated drug due to the acidic pH within the intracellular organelles of the cancer cell line. Cell viability tests in the lab verified all forms of Dox in the study to have comparable cytotoxicity to cause cancer cell death in vitro.

Inspired by in-lab pharmacokinetics and tumor accumulation studies, the research team tested the anti-tumor effects of Dox-CBD-SA in vivo in tumor-bearing mice. For this, they injected the MMTV-PyMT orthotopic tumor-bearing mice with single intravenous injections of several Dox forms via the tail vein. The work showed that pre-conjugation of Dox with SA provided a higher therapeutic effect than the conjugation of doxorubicin with endogenous SA in situ. Importantly, Dox-CBD-SA showed greater therapeutic potential compared to Dox-SA by extending the survival rate and inducing tumor remission in 2 of 12 mice. The data supported the superiority of the Dox carrier compared to the unmodified SA, relative to antitumor efficacy.



TOP: Dox-CBD-SA treatment shows reduced toxicity. Aldoxorubicin or Dox-CBD-SA (20 mg/kg on a Dox basis) was administered to tumor-free FVB mice via tail vein injection on day 0. (A to D) Plasma cytokine concentrations on day 3. (E) Red blood cell (RBC) counts on day 6. (F) White blood cell (WBC) counts on day 3. (G) Spleen weights on day 16. Data represent mean  $\pm$  SEM. Two experimental replicates. Statistical analyses were done using ANOVA with Tukey's test. \*P

Dox also induced [immunogenic cell death](#) (ICD) to stimulate [tumor-infiltrating lymphocytes](#) (TIL), which were biomarkers for [favorable prognosis](#) in multiple cancers. Sasaki et al. therefore analyzed TILs after treatment with the new drug in the present work and recorded enhanced filtration of lymphocytes for antitumor effects. The researchers observed lower toxicity of Dox-CBD-SA compared to doxorubicin due to its slow release under physiological pH. For instance, doxorubicin increased the plasma concentration of inflammatory cytokines and decreased the red blood cell counts, white blood cell counts and hemoglobin concentration. By contrast, the adverse impact of Dox-CBD-SA was mild. The work suggested a reduced toxicity spectrum during treatment after pre-conjugation of Dox with CBD-SA.

The research team further investigated if combining Dox-CBD-SA with CPI (checkpoint inhibitors) showed greater therapeutic effects compared to doxorubicin therapy with CPI. For this, they used the [MC38 colon carcinoma model](#), which was [immunogenic](#) but not curable with [Dox monotherapy](#) alone. They noted the new drug induced immunogenic cell death synergistically with the clinical CPI to achieve superior antitumor effects compared to doxorubicin and a clinical CPI.

In this way, Koichi Sasaki and colleagues developed a new therapeutic agent Dox-CBD-SA, which accumulated in tumors to activate host antitumor immunity. Monotherapy of the new agent suppressed orthotopic MMTV-PyMT breast tumor growth in an animal model and prolonged their survival. When combined with immune checkpoint inhibition, the drug completely eradicated tumors in an immunogenic MC38 mouse model. They conclude that the pre-conjugation of CBD-SA may hold potential for clinical translation during cancer therapy as an antitumor drug carrier

**More information:** Koichi Sasaki et al. Engineered collagen-binding serum albumin as a drug conjugate carrier for cancer therapy, *Science Advances* (2019). [DOI: 10.1126/sciadv.aaw6081](https://doi.org/10.1126/sciadv.aaw6081)

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