

Noninvasive monitoring of chronic kidney disease (CKD) using pH and perfusion imaging

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Representative pixel-by-pixel B0 inhomogeneity calculation (2B0) maps for RD (regular diet) and HP (high protein) diet mice. 2B0 are generated using WASSR (water saturation shift referencing) experiment for MRI data acquisition



performed using 0.5 PPRF (radiofrequency) saturation power and 42 offsets between +1.5 ppm and -1.5 ppm. The mean frequency shift in water frequency and the standard deviation is mentioned in the image. (A), (B) Control (Mut+/-) and mutant (Mut-/-) mice fed on a regular diet. (C), (D) Mut+/- and Mut-/- mice fed on a high protein diet. Credit: Science Advances, doi: 10.1126/sciadv.aaw8357

Chronic kidney disease (CKD) is a feature of inherited metabolic disorders such as <u>methylmalonic acidimia</u> (MMA), which can cause impaired growth and low metabolic activity to delay the diagnosis and management of renal disease. In a new report now published on *Science Advances*, KowsalyaDevi Pavuluri and colleagues at the interdisciplinary departments of Radiology and Radiological Science, Functional Brain Imaging, Metabolic Genetics and Pediatrics in the U.S., designed an alternative strategy to monitor renal function. The research team based the protocol on administering a pH sensitive MRI agent to assess disease progression in a mouse model of MMA. The study successfully facilitated the use of MRI for early detection and monitoring of CKD.

Organic acidemias are a group of rare inborn metabolic errors caused by the disturbances of amino acid metabolism that result in large, abnormal accumulations of toxic organic acids in tissues and fluids—including urine. Collectively, this group of inborn errors of metabolism can cause notable morbidity and mortality in infancy and childhood. As a result, these disorders are now routinely screened in newborn panels for early diagnosis and treatment. Scientists have established gene knockout and tissue-specific transgenic mouse models of MMA in research studies to resemble the human disease, due to its severity and ubiquity. Although early identification and treatment has improved the life expectancy and clinical course of MMA, <u>patients remain at risk</u> of chronic complications including CKD and renal failure.



Renal function can be determined using the standard index of <u>glomerular</u> filtration rate (GFR), since the <u>direct measurement of GFR</u> (mGFR) is invasive and cumbersome. As an alternative to mGFR, clinicians also routinely estimate GFR (eGFR) using <u>creatinine and/or cystatin-C</u>. However, the routine computation of serum biomarkers of renal disease is substantially altered in severely <u>growth impaired and protein-restricted</u> patients, while eGFR is limited to patients with <u>asymptomatic renal</u> disease. The development of alternative methods to detect early stage CKD while facilitating the treatment of associated comorbidities are still evolving, the techniques must also enable timely planning for renoprotection or renal replacement therapies, including transplantation to specific patient populations.





In vitro calibration plots and pH maps using iopamidol in human blood serum. (A) Structure of iopamidol with exchangeable protons highlighted, which produce CEST contrast at 4.2 and 5.5 ppm at a relative concentration of 2:1. (B) CEST Z-spectra of iopamidol in blood serum at $\omega 1 = 4 \mu T$ for pH = 5.3, 6.1, 6.5, 6.9, and 7.3. (C) Calibration plot used to calculate in vitro and in vivo pH; variation of experimental ST ratio at different pH values was given by pH = p1 × (ST ratio)3 + p2 × (ST ratio)2 + p3 × (ST ratio)1 + p4, with p1 = -0.01174, p2 = 0.1653, p3 = -0.927, and p4 = 7.598. The root mean square error of the fit was 0.0714 for pH values from 5.5 to 7.3. (D) pH maps of iopamidol-serum phantom. In the map, pH values below 5.5 were determined inaccurately. (E) Error bar plot representing the accuracy in MRI pH measurements compared to that of electrode pH for iopamidol-serum phantom. Error bars (blue) were obtained by calculating the mean SD in pH over a region of interest (ROI) drawn enclosing the entire tube in the phantom. Science Advances, doi: 10.1126/sciadv.aaw8357

Research teams are therefore keen to develop new tools to detect renal disease to meet the needs of MMA and CKD populations. For instance, magnetic resonance imaging (MRI) can produce high resolution images with exquisite soft tissue contrast to <u>detect renal pathologies</u>. Researchers have also <u>impressively measured functional information</u> on kidney clearance or the count of glomeruli using MR contrast agents during <u>renography</u>. To create pH images most researchers have used <u>chemical exchange saturation transfer (CEST) MRI</u> as a premier technology. The technology can detect low concentrations of contrast agent by applying saturation pulses on labile (unstable) protons to destroy their magnetization and transfer the signal loss to water via chemical exchange. The resulting chemical shift dependence is an important feature able to discriminate between agents of <u>multicolor</u> or <u>multifrequency MRI</u>. For pH imaging studies, <u>iopamidol is an excellent pH probe</u> used and substantiated in previous studies.



In the present work, Pavuluri et al. used CEST MRI to functionally image kidneys in an MMA mouse model and investigate the progression to CKD. They designed a time-efficient CEST MRI protocol to study a nonionic contrast agent known as <u>iopamidol</u> to yield perfusion and pH maps of the kidneys. The research team envision that the new method will determine mGFR (direct measurements of GFR) as a useful technique to diagnose and monitor CKD. The team first tested the pH sensitivity of iopamidol and assessed the pH mapping protocols across a range of pH values, followed by testing several radiofrequency (RF) saturation powers best suited for blood serum. Using the multifrequency pH mapping protocol developed in the work, the researchers measured pH values between 7.3 and 5.7, suited for live imaging animal studies.





Contrast agent uptake for the four different groups of MMA mice. (A) Average iopamidol uptake with time for RD Mut+/– and RD Mut–/– mice. RD Mut+/– mice display higher uptake of contrast agent [>25% (n = 5)] than RD Mut–/– mice [>12% (n = 4)]. (B) Average iopamidol uptake with time for HP Mut+/– and HP Mut–/– mice. Similar to RD Mut+/– mice, >25% contrast was observed for HP Mut+/– mice. HP Mut–/– mice displayed the lowest contrast [~4% (n = 3)], indicating the lowest iopamidol uptake for these mice with more advanced kidney disease. (C and D) Corresponding time-averaged maximum contrast images of RD Mut+/– and RD Mut–/– mice. (E and F) Maximum contrast images of HP Mut+/– and HP Mut–/– mice. Science Advances, doi: 10.1126/sciadv.aaw8357

Subsequently, they developed an in vivo CEST MRI protocol to study a <u>mouse model of MMA</u> carrying a mutation $(Mut^{-/-})$ and displaying varying degrees of kidney disease within four groups alongside control littermates $(Mut^{+/-})$. Preceding research work had shown that exposure to a high protein diet caused a massive elevation of biomarkers related to CKD in the mutant animals. The research team administered iopamidol to the animals and acquired CEST MRI data and observed the strongest contrast in the control mice and the weakest contrast in the mutant mice on a high protein diet. The results on iopamidol uptake could thus differentiate between groups during early disease progression.





In vivo filtration fraction (FF) results for MMA mice. (A) FF calculated by taking the time-averaged FF for 10 images. The FF is defined as the percentage of kidney pixels with >20% of maximum contrast, and the SD in FF is calculated by comparing the 10 time points used. RD and HP Mut+/– controls display FF of ~100% (n = 5), whereas HP Mut-/– mice show the lowest FF, indicating that FF could be a CEST MRI metric for detecting renal disease. The blue, green, and red colors in the perfusion maps are used to denote the Mut+/–, RD Mut-/–, and HP Mut-/– mice, respectively. The decrease in FF was in accordance with a decrease in weight of the mice, which is a secondary indicator of disease progression for these mice. A linear correlation was observed between weights of mice and FF with R = 0.86 (for all groups of mice) and R = 0.9 (only considering the RD and HP Mut-/– mice). (B and C) Time-averaged FF images of RD mice. (D and E) Time-averaged FF images of HP mice. RD Mut-/– mice (n = 4) have lower FF than RD Mut+/– controls but have higher FF than HP Mut-/– mice (n = 3). Science Advances, doi: 10.1126/sciadv.aaw8357

The research team compared the perfusion results and filtration fraction (FF; metric of perfusion) across all groups of MMA mice by test imaging with iopamidol to detect kidney disease. Control animal models on the regular diet and high protein diet displayed FF > 98 percent, whereas animal models with moderate kidney disease displayed a



moderate reduction in iopamidol perfusion (FF, ~79 percent). Furthermore, animal models with severe kidney disease displayed a substantial reduction in iopamidol perfusion (FF, ~50 percent).

To establish the suitability of the MRI protocol, Pavuluri et al. obtained pH maps using the CEST MRI data for the four groups of mice. Regular diet and high protein diet fed control mice displayed homogenous pH values of 6.50 across the entire kidney slice imaged. In contrast mutant mice on a regular diet displayed a lower average pH (~6.1) and mutant mice on a high protein diet showed slightly lower pH (~6.0). The in vivo imaging data were also consistent with the blood work and weights of the animals obtained in the study.





In vivo pH results of two groups of mice maintained on RD and HP diets. (A) pH histograms calculated for two representative RD Mut+/- and two representative RD Mut-/- mice. For Mut+/- mice (n = 5), more than 80% of the detected pixels display pH = 6.50, whereas for RD Mut-/- mice (n = 4), an acidic pH of 6.05 to 6.32 was observed for >20% of the kidney pixels. (B and C) Time-averaged pH maps of these RD Mut+/- and RD Mut-/- mice. The average pH dropped to 6.05 to 6.32 for RD Mut-/- compared to 6.50 for RD Mut+/– controls. The pH ranges of RD Mut-/– mice were ± 0.06 and ± 0.09 pH units compared to ± 0.02 and ± 0.01 pH units for RD Mut+/– mice. (D) pH histograms calculated for two representative HP Mut+/- mice and two representative HP Mut-/- mice. For HP Mut+/- mice, more than 80% of the detected pixels display a mean pH of 6.50, whereas for HP Mut-/- mice (n = 3), an acidic mean pH of 6.10 to 5.83 was observed. (E and F) Time-averaged pH images of Mut+/-and Mut-/- controls of HP mice. pH was further lowered to 5.83 for the most severely diseased mice. The pH was distributed over a narrow range of 6.50 ± 0.02 for both RD and HP Mut+/– mice, while this range significantly increased to ± 0.30 and ± 0.46 along with a decrease in mean pH for HP Mut-/- mice. Science Advances, doi: 10.1126/sciadv.aaw8357

In this way, KowsalyaDevi Pavuluri and co-workers developed a new CEST MRI protocol based on iopamidol administration to test perfusion and detect pH changes in mice exhibiting different stages of renal function. The research team used filtration fraction (FF) as the metric of perfusion to estimate isopamidol perfusion through the kidneys due to minimal modeling required to measure it. Based on the results, Pavuluri et al. determined the new CEST MRI protocol as a promising technique to assess disease progression in time. The method facilitated treatment responses in animal models of kidney disease, with potential for immediate translation in patients with disorders similar to MMA.

More information: Devi Pavuluri et al. Noninvasive monitoring of



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