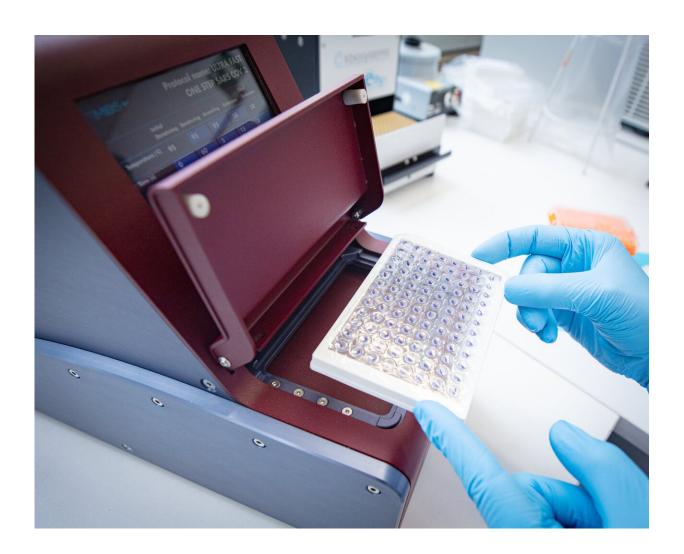


New method performs coronavirus test 10 times faster

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Whether a person is infected with SARS-CoV-2 can be detected in tests. The genetic material is multiplied in a thermal cycler. The thermal cycler used in the new study (see photo) enables around 570 analyses per hour. Credit: Bielefeld University/M.-D. Müller



Currently, it takes more than two hours to perform and evaluate a test for SARS-CoV-2—meaning that a laboratory can test only a very limited number of people per day. In a study with several cooperation partners, cell biologists at Bielefeld University have now developed a method that delivers results around 10 times faster. "The test takes only about 16 minutes," says Professor Dr. Christian Kaltschmidt from Bielefeld University's cell biology research group. "The method is also cheaper than conventional tests."

More than 10 million <u>coronavirus</u> infections have now been confirmed worldwide. An effective vaccine or therapy for SARS-CoV-2 is not yet available. On top of this: not every infected person develops symptoms. Therefore, testing is currently the most effective method of curbing the spread of the virus: those who are infected are then isolated and no longer spread the virus.

The most common procedure for testing whether someone has become infected with SARS-CoV-2 is a so-called PCR test. These use the genetic material of the virus as a basis. This is also what the Bielefeld scientists did in their study. PCR tests always follow a similar pattern. First, genetic material is needed from a test person. This is usually obtained by a smear test in the mouth, nose, or throat. "If a person is infected with SARS-CoV-2, the sample will also contain genetic material from the virus in the form of what is called RNA," said Kaltschmidt. The RNA molecules are isolated in a chemical process. However, the sample does not contain enough RNA for a test to detect it immediately. Therefore, it has to be amplified.

This is done with a so-called <u>polymerase chain reaction</u>—which is what gave the PCR method its name (polymerase chain reaction). It is carried out in a device called a thermal cycler. This raises and lowers the



temperature according to a predetermined program. In the presence of certain additives, an enzyme with a copy function, and stability at high temperatures, the <u>genetic material</u> multiplies until there is enough to detect SARS-CoV-2—if someone is infected.

The Bielefeld researchers used a special thermal cycler—the NEXTGENPCR—for their process. Thanks to the special design including several temperature zones, reactions in the device run particularly effectively and fully automatically. "We have oriented the procedure towards the so-called Drosten protocol of the Berlin Charité and the protocol of the Center of Disease Control and Prevention in Atlanta," says Kaltschmidt. These are standards for SARS-CoV-2 tests. Using their method, the researchers were able to repeat the results of conventional PCR tests—only in a much shorter time and with less effort.

Special device can analyze 570 tests per hour

The thermocycler was developed by the Dutch company Molecular Biology Systems B.V. For the coronavirus tests, the developers wrote software that reduces both the time required and the number of steps involved. "We received a lot of positive feedback," said Gert de Vos, founder and CEO of Molecular Biology Systems. The device can analyze several samples in parallel—this means that a single thermal cycler can be used to analyze around 570 samples per hour. Molecular Biology Systems is now working with governments and private laboratories in Africa, Europe, the Middle East, and the U.S..

Kaltschmidt sees many advantages in the new process. For example, such a test could be used especially where rapid results are required. "When <u>cruise ships</u> resume operations, for example, they could test each person in a short time before they board the ship."



More information: Ehsan Asghari et al. Ultra-fast one-step RT-PCR protocol for the detection of SARS-CoV-2, (2020). DOI: 10.1101/2020.06.25.20137398

Provided by Bielefeld University

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