

# Self-collected saliva samples prove effective for diagnosing COVID-19

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Credit: Unsplash/CC0 Public Domain

Researchers at Memorial Sloan Kettering Cancer Center (MSK) have found that SARS-CoV-2 genetic material can be reliably detected in self-collected saliva samples at a rate similar to that of nasopharyngeal and

oropharyngeal swabs. The rate of detection using saliva samples was similar across different testing platforms, and saliva samples remained stable for up to 24 hours when stored with ice packs or at room temperature, according to a new study in *The Journal of Molecular Diagnostics*. Oral rinses, which have been suggested as another alternative to nasal swab collection, did not reliably diagnose COVID-19.

"The current pandemic has placed a significant strain on the supply chain, from swabs to the [personal protective equipment](#) (PPE) [healthcare workers](#) need to safely collect samples," explained lead investigator Esther Babady, Ph.D., D(ABMM), FIDSA, director of the Clinical Microbiology Service at Memorial Sloan Kettering Cancer Center, New York, NY, U.S.. "The use of self-collected [saliva](#) has the potential to minimize healthcare worker exposure and decrease the need for specialized collection devices, such as swabs and viral transport media."

The study was conducted at MSK in New York City at the peak of the regional outbreak between April 4 and May 11, 2020. Study participants were 285 MSK employees who needed to be tested for COVID-19 because they had symptoms of the virus or had been exposed to someone who had the virus. Each participant provided paired samples: a nasopharyngeal swab and oral rinse; a nasopharyngeal swab and a saliva [sample](#); or an oropharyngeal swab and a saliva sample. All samples to be tested were stored at [room temperature](#) and transported to the laboratory within two hours.

**SALIVA COLLECTION**

**PURPOSE**

Collecting saliva samples for COVID-19 Qualitative RNA PCR.

**COLLECTION KIT**

50 mL conical tube

**DIRECTIONS**

1. Swallow and bring up saliva from the back of your throat
2. Spit at least **3.0 mL** of saliva into the empty conical tube (Completely fill the lower conical portion with saliva. If you notice excess bubbles, cap tube and tap down to ensure the saliva fills the entire conical portion of the vial)
3. Close tube and **tightly** secure the cap
4. Apply barcoded specimen label.

Place in specimen bag and deliver to your local laboratory.

5/19/2020

Instructions given to individuals when collecting their saliva sample. Credit: Memorial Sloan Kettering Cancer Center

The agreement between the saliva test and the oropharyngeal swab was 93 percent, with a sensitivity of 96.7 percent. In comparison with the nasopharyngeal swab, the agreement of the [saliva test](#) was 97.7 percent, with a sensitivity of 94.1 percent. Oral rinses were only 63 percent effective in detecting the virus, with an overall agreement with nasopharyngeal swab of only 85.7 percent. To test for stability, [saliva samples](#) and nasopharyngeal samples with a range of viral loads were stored in a transport cooler at 4° C or at room temperature. No significant difference in virus concentration was detected in any samples at the time of collection, eight hours later, and 24 hours later. These results were validated on two commercial SARS-CoV-2 PCR platforms, and overall agreement between the different testing platforms was over 90 percent.

Dr. Babady noted that validation of sample self-collection methods holds great promise for broad testing strategies that would mitigate infection risk and PPE resource utilization. "The current 'test, track, and trace' public health approach to surveillance relies heavily on testing for both diagnosis and surveillance," she stated. "The use of self-collected saliva provides a cheaper and less invasive option for viable sample collection. It's certainly easier to spit in a cup twice a week than undergoing frequent nasopharyngeal swabs. This can improve patient compliance and satisfaction particularly for surveillance testing, which requires frequent sample collection. Since we also showed that the virus was stable at room temperature for at least 24 hours, saliva collection has potential for use at home."

**More information:** N. Esther Babady et al, Performance of Severe Acute Respiratory Syndrome Coronavirus 2 Real-Time RT-PCR Tests on Oral Rinses and Saliva Samples, *The Journal of Molecular Diagnostics* (2020). [DOI: 10.1016/j.jmoldx.2020.10.018](https://doi.org/10.1016/j.jmoldx.2020.10.018)

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