

An illuminated 3D tour of lower back pain

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Fluorescently labeled molecules target unfurled collagen proteins to create detailed images and 3D maps of damaged soft tissues in the spines of mice and rats. Credit: American Chemical Society.

Back pain affects many people at some point in their lives, and a common cause is damage to the squishy discs or flexible, rubbery tissues of the spine. However, observing this damage at an early stage is difficult with current imaging methods. Now, researchers reporting in *ACS Nano* can see microscopic soft tissue destruction in animal spines



by targeting denatured collagen with fluorescent molecules.

Anywhere along the spine, from the neck to tail bone, can become uncomfortable when its soft and protective tissues, including the cartilage and jelly-like intervertebral discs, become damaged and lose their structure. Daily wear-and-tear, as well as some disorders, such as facet joint osteoarthritis or ankylosing spondylitis, can degrade and unfurl the collagen proteins that give these tissues their bounce and flexibility. Detecting compromised collagen early could help patients get relief before the pain becomes severe, but this is very difficult to do with existing medical technologies, such as X-rays and magnetic resonance imaging (MRI). Previously, Yang Li and colleagues developed a collagen hybridizing peptide (CHP) probe that specifically binds unfurled collagen molecules, which happens when they deteriorate and lose their ability to cushion vertebrae. So, Li, Kuibo Zhang, Hong Shan and colleagues wanted to test if CHP labeled with fluorescent tags could be used as an imaging method to identify collagen destruction in the body.

To make the peptide probe more stable in the body, the researchers modified CHP by substituting a hydroxyl group with fluorine and then attaching a fluorescent dye to it. When healthy mice and rats were injected with the fluorescent dye-labeled CHP and imaged with nearinfrared fluorescence (NIRF), the team could confirm that the fluorescing molecules accumulated on the soft tissues between the vertebrae. Then the researchers removed a portion of the animals' spines and imaged them with light sheet fluorescence microscopy. This technique produced precise 3-D maps, which revealed denatured collagen.

Because CHP is known to specifically target damaged collagen, the team says their imaging experiments show that even healthy animals can have a modest degree of deteriorated collagen around load-bearing joints, especially in the lower back. In additional experiments, both the NIRF



images and 3-D maps generated with the new method detected <u>collagen</u> deterioration in animal models of spinal injury before structural changes were visible in tissues on MRI scans. Finally, the researchers applied dyelabeled CHP as a stain to intervertebral disc slides from people that had undergone spinal surgeries. The fluorescence intensity of the stain rose substantially as the level of disc degeneration increased. Based on these results, the researchers say that their molecular-level technique could be developed in clinical studies for earlier diagnosis and targeted therapeutic treatments for patients with back pain.

More information: Lei Liu et al, Molecular Imaging of Collagen Destruction of the Spine, *ACS Nano* (2021). <u>DOI:</u> <u>10.1021/acsnano.1c07112</u>

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