

Immunological memory provides long-term protection against coronavirus

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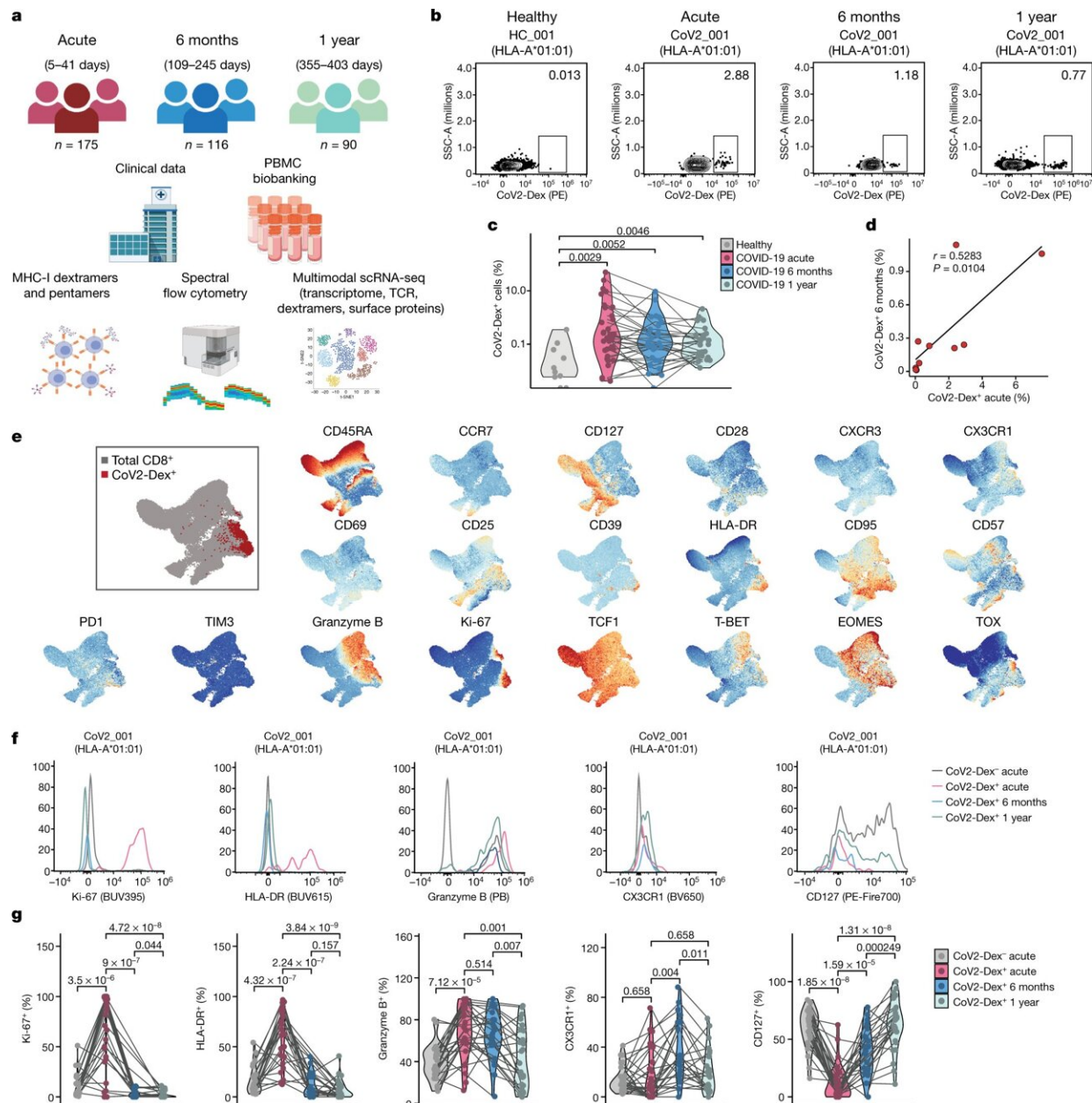


Fig. 1: Characteristics of antigen-specific CD8⁺ T cells during acute and memory phases of SARS-CoV-2 infection. a, Overview of study design. PBMC, peripheral blood mononuclear cell. b, Representative plots of CoV2-Dex staining. PE, Phycoerythrin. Numbers in the plots indicate percentage of parent population. c, Frequency of CoV2-Dex⁺ cells in healthy donors and patients with COVID-19 during acute infection and 6 months and 1 year after infection. Each dot represents an independent donor at the indicated timepoint (n = 10 healthy, n = 37 acute, n = 32 6 months, n = 29 1 year after infection). P values are shown. d, Linear regression of frequency of CoV2-Dex⁺ cells 6 months after infection as a function of CoV2-Dex⁺ cell frequencies during acute infection (n = 11). The P value was calculated with t-statistic. e, Uniform manifold approximation and projection (UMAP) plots of marker expression for up to 2,000 CD8⁺ T cells from each sample collected during acute infection (n = 37) analyzed by spectral flow cytometry. Regions with high marker expression appear in red. An overlay of CoV2-Dex⁺ cells (red) and total CD8⁺ T cells (gray) is shown in the top left. f, Representative histograms showing expression of selected markers on CoV2-Dex[−] and CoV2-Dex⁺ cells. g, Frequency of Ki-67⁺, HLA-DR⁺, granzyme B⁺, CX3CR1⁺ and CD127⁺ cells in CoV2-Dex[−] (gray) and CoV2-Dex⁺ cells during acute infection and 6 months and 1 year after infection. Analysis was conducted on paired samples from acute infection versus 6 months and/or 1 year after infection (n = 28 acute, n = 24 6 months, n = 29 1 year). The gray lines connect individual donors sampled at different timepoints. P values were calculated using a Wilcoxon–Mann–Whitney test in c and g and corrected for multiple comparisons in g. All tests were performed two-sided. Credit: DOI: 10.1038/s41586-021-04280-x

Many questions about how exposure to SARS-CoV-2 by infection or immunization might result in long-term protective immunity remain unanswered. Onur Boyman, head of the Department of Immunology, and his research team at the University of Zurich and the University Hospital Zurich, have now taken a closer look at how this long-lived protection is formed. Together with researchers from ETH Zurich, they

identified specific signaling pathways that determine when immune cells develop into so-called memory T cells.

From short-lived killers to long-term memory T cells

Virus-specific antibodies produced by B [cells](#) are insufficient to effectively protect against the novel coronavirus. The cellular immune response to SARS-CoV-2 is just as important. Here, virus-specific CD8+ T cells play a crucial role, as they can identify and kill the cells that have been infected by the virus. These cytotoxic T cells eliminate viruses that are hidden inside the host cells and help prevent the spread of millions of newly formed viruses. "These T cells are usually active for only a short time and disappear quickly. When it comes to establishing long-term protective immunity, it is important to generate long-lived memory T cells that are activated very quickly upon re-exposure to the virus," explains Onur Boyman. This latter ability is referred to as immunological memory.

Previous studies have focused on the whole CD8+ T cell population that formed in response to the virus. Boyman and his team have now succeeded in tracking individual clones of SARS-CoV-2-specific CD8+ T cells in patients with COVID-19, from the acute viral [infection](#) up to one year after recovery. The researchers were also able to identify the signaling pathways responsible for the transition of CD8+ T cells from short-lived killers to long-lived memory cells—and they found a distinct molecular signature.

Immune messengers determine the cell type

In their study, the researchers were able to demonstrate that the signature of long-lived memory CD8+ T cells was already present during acute SARS-CoV-2 infection, and these cells could thus be distinguished from

their short-lived counterparts at an early stage. "The distinct signature of memory cells contained signals of immune messengers, such as interferons, which are an important part of the immune response against SARS-CoV-2 and also contribute to controlling viral infections," says Onur Boyman.

Immune response varies from one patient to another

The study reveals the complex way in which immunological memory to SARS-CoV-2 is—or is not—formed and maintained. While some infections result in robust and long-lasting T cell [memory](#), others fail to do so. The newly identified signature makes it possible to determine which type of infection—e.g., mild or severe, systemic or limited to mucosal membranes—gives rise to sustained immunity. The immune response is also shaped by vaccines, which contain different ingredients and adjuvants. "While everyone responds differently to the virus or a vaccine, cellular immunity plays a crucial role in preventing severe cases of COVID-19 in both vaccinated and recovered people," says Boyman.

More information: Sarah Adamo et al, Signature of long-lived memory CD8+ T cells in acute SARS-CoV-2 infection, *Nature* (2021). [DOI: 10.1038/s41586-021-04280-x](https://doi.org/10.1038/s41586-021-04280-x)

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