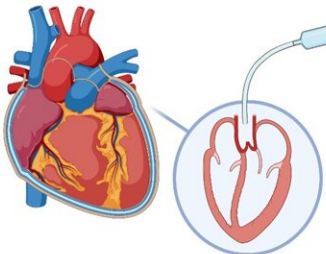


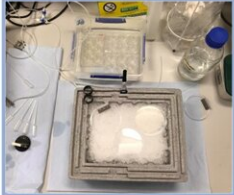
Lab-grown pig heart tissue could help replace live animals in heart disease research

March 8 2022

Heart collection and setup



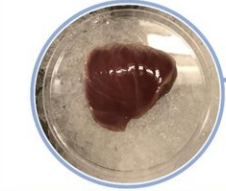

1 Remove the heart maintaining the pericardial membrane intact and slowly inject 100 ml of ice-cold cardioplegia solution through the aorta




2 Tissue dissection and sample preparation equipment:
 -Polystyrene box full of ice
 -Petri dish
 -Agarose cushion prepared into Petri dish lid
 -Sterile forceps and scalpel

Tissue block preparation

3 Quickly separate the ventricles from the rest of the heart by cutting along the left anterior descending artery and the posterior descending artery

4 Dissect 8x8 mm tissue blocks by making incisions through the full thickness of the ventricular wall with a single edge steel blade



Tissue block embedding

5 Gently place each tissue block on the agarose cushion with the epicardium facing down onto the agarose.




6 Place the 3D printed plastic ring around the tissue block and pour the low melting agarose solution into the plastic ring until all the tissue is covered and the solution reaches over the edge of the ring



7 Once solidified cut off the agarose excess along the ring top then remove the resulting block from the ring. Square one side of the agarose cylinder to obtain a flat surface (red dotted line) and to orientate the tissue on the vibratome specimen holder facilitating the alignment of the blade



Critical point: enables the epicardial surface to adhere thoroughly to the agarose to flatten the cutting surface

Slices cutting and culture

8 Position the vibratome's blade on top of the block and assign as starting point. Set the cutting thickness to 400-500 μm and start slicing at 0.03 mm/s speed and 1 mm amplitude



9 Once cut, collect each slice and transfer in the recovery bath for at least 30 minutes to up to 4-5 hours



10 Slices can be cultured efficiently in the pillared Petri dish, changing medium every 24 hours (static) or using BioFlo120 bioprocessor which pumps the medium from a reservoir to the culture dish via a custom 3D-printed insert (dynamic)




11 Slices viability, cellular composition and epicardial cells morphology can be assessed by live staining and/or histological analysis after fixation.



Fig. 1: Porcine epicardial slices preparation. Schematic representation of porcine ventricles sampling and slicing. Heart collection and setup: (1) Heart is harvested maintaining the pericardium and retroperfused with ice-cold cardioplegia solution. Ventricles are then separated from the heart avoiding touching the

epicardium, to prevent damage. (2) Embedding area must be set up prior to tissue slicing to allow all components to cool down to ice-cold temperature. Tissue block preparation: (3) Cardiac tissue is placed on the ice-cold Petri dish and (4) blocks of tissue 8 mm ×8 mm are dissected with a single edge steel blade. Tissue block embedding: (5) The epicardial side of the slice is flattened onto a cold agarose surface and placed in the middle of a custom-made 3D printed ring. 5–8 ml of 30–37 °C low melting agarose is poured to embed the tissue block. (6) Once solidified, the block is removed from the ring and (7) squared on one side to allow a better alignment with the vibratome's blade (red dashed line). Slice cutting and culture: (8) Embedded tissue blocks are oriented along the squared side and glued to the vibratome specimen holder. The blade is aligned to the epicardial surface and 400–500 µm thick slices are cut using a high precision vibratome. (9) Slices are allowed to recover for at least 30 min in a recovery bath. (10) Epicardial slices are cultured epicardium-up on 8-mm-high pillars cast at the bottom of a Petri dish. (11) En face immunohistochemistry of WT1 (red) and mesothelin (MSLN, green) shows the presence of a continuous epicardial layer, characterized by typical morphology and marker expression (WT1+ cells indicated with full arrowhead, WT1– MSLN+ cells indicated with empty arrowhead). This figure was created using BioRender.com. Credit: *npj Regenerative Medicine* (2022). DOI: 10.1038/s41536-021-00202-7

A new way to replicate what happens inside the heart after cardiac arrest could open new avenues for the study of heart regeneration whilst reducing the use of live animals in research, according to a study from the University of Surrey and King's College London.

Researchers have developed a process that involves obtaining and growing thin slices of pig [heart](#) tissue which include both the epicardium—the most external layer of the heart that contains cells that can promote its recovery—and underlying heart muscle.

The team treated the epicardial slices with stimulating compounds, showing that cells become activated in a way that replicates what

happens in the heart after a heart attack. The new process was able to reproduce observations typically obtained in live animal models.

Dr. Paola Campagnolo, lead author of the study and senior lecturer in molecular cardiovascular sciences at the University of Surrey, said: "This research typifies the One Health, One Medicine ethos at the University of Surrey, as our [model](#) could help us understand how to stimulate the repair process after heart attacks without the need to use live animals in the research. We are hopeful our model could lead to better health outcomes for humans and reduce the reliance on animal experiments in cardiovascular science."

According to the British Heart Foundation, there are around 7.6 million people living with heart or circulatory disease in the UK. This disease causes a quarter of all deaths in the UK.

The ability of the heart to recover after an injury is severely limited by the low number of regenerating cells within its tissue. The current research models and strategies aimed at improving the heart's repair process are mainly based on surgical procedures performed on laboratory animals.

Dr. Davide Maselli, postdoctoral research associate and first author of the paper, said: "This work provides an innovative tool to study the healing from a [heart attack](#). We believe that our model could be useful to dissect the role of different cells in the reparative process. In our consideration, it is extremely important that every step forward in this field delivers a clinical perspective for the patients while reducing the burden on research animals."

The research, published in the journal *npj Regenerative Medicine*, proposes a system to study the regeneration of the heart in a laboratory dish and could therefore lead to a reduction in the number of small

[animals](#) used in cardiovascular research.

More information: D. Maselli et al, Epicardial slices: an innovative 3D organotypic model to study epicardial cell physiology and activation, *npj Regenerative Medicine* (2022). [DOI: 10.1038/s41536-021-00202-7](https://doi.org/10.1038/s41536-021-00202-7)

Provided by University of Surrey

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