

Gene-delivering viruses reach the brain in step toward gene therapy for neurological diseases

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Gene therapies can treat, even potentially cure, certain genetic diseases, but it is challenging to deliver the treatments to the parts of the body where they are needed. Researchers have engineered viruses called adeno-associated viruses (AAVs) to deliver cargo—such as a functioning copy of a gene—to specific cells and organs, but they don't always get to their desired destination.

Researchers at the Broad Institute of MIT and Harvard have now developed a family of AAVs that is able to reach a particularly



challenging target tissue—the brain. The team shows, in a study published in *Med*, that their AAVs are more than three times better at delivering their cargo into the <u>primate brain</u> than the current leading AAV delivery vehicle, AAV9.

The new AAVs can cross the blood-brain barrier, which keeps many drugs from getting into the brain. They also accumulate much less in the liver than AAV9, potentially reducing the risk of liver side effects that have been seen in other AAV9-based gene therapies. This family of AAVs, called the PAL family, could be a safer and more efficient way to deliver gene therapies to the brain.

The AAVs were designed in the lab of Pardis Sabeti, who is an institute member at the Broad, a professor at Harvard University and the Harvard T.H. Chan School of Public Health, and a Howard Hughes Medical Institute investigator.

"We generated a massive pool of randomly generated AAV capsids and from there narrowed down to ones able to get into the brain of both mice and macaques, deliver genetic cargo, and actually transcribe it into mRNA," said study lead author Allie Stanton, a Harvard Medical School graduate student in the Sabeti lab.

A protective shell

Gene therapies consist of DNA, RNA, or other molecules that are transported around the body by delivery vehicles, or vectors. AAVs are promising vectors because as viruses, they are effective at delivering their contents into cells. Scientists replace the AAVs' natural payloads with therapeutic DNA, gene-editing machinery, or other genetic information that they want to get inside cells to treat disease.

"AAVs are a really good gene therapy vector because you can put



whatever you want inside of its shell, which will protect it and get it into a wide variety of cell types," said Stanton.

However, the majority of an injected AAV dose typically ends up in the liver, meaning that high doses of AAV are required to get even a fraction into a different target tissue, such as the brain. In some cases, these high doses have resulted in <u>liver damage</u> and even death in clinical trials.

Engineering vectors to efficiently target specific cells or organs could help reduce these unwanted side effects. Gene therapy researchers are working to make AAVs safer and more effective by changing the amino acid makeup of the virus' shell, or capsid. Because there are billions of possible synthetic AAV capsids, scientists can modify thousands to millions of viruses at a time to search for ones that fit a specific purpose—such as crossing the blood-brain barrier.

Building better vectors

To develop a delivery system that might one day be used for hard-totreat neurological diseases, Stanton and colleagues focused on pinning down AAVs that cross the blood-brain barrier. They turned to a method developed in the Sabeti lab called DELIVER, in which scientists generate millions of capsids and look for AAVs that successfully deliver their payload to certain target cells. Using DELIVER, the team developed the PAL family of AAVs that cross the <u>blood-brain barrier</u> more effectively than AAV9—the only FDA-approved viral vector for use in the <u>nervous system</u>.

They found that the PAL AAVs were three times more effective at producing therapeutic mRNA in the macaque brain compared to AAV9.

The team also found that the engineered viruses had a unique pull to the



brain. PAL-treated macaques had one-fourth of the viral material in their livers as AAV9-treated primates did, suggesting that the new capsids could help limit the liver toxicity of other <u>gene therapies</u>.

The authors say PAL AAVs could potentially work in humans given how similar macaques are to humans, but added that the AAVs didn't work well in mice, making it difficult to test these vectors in mouse models of disease. Moving forward, the team hopes that their work will provide a starting point for even more effective viral vectors.

"We are encouraged by the early results of the PAL family AAVS, and can see several promising lines of investigation using directed evolution and engineering to further increase their efficiency," Sabeti said.

More information: Alexandra C. Stanton, Systemic administration of novel engineered AAV capsids facilitates enhanced transgene expression in the macaque CNS., *Med* (2022). <u>DOI: 10.1016/j.medj.2022.11.002</u>. <u>www.cell.com/med/fulltext/S2666-6340(22)00456-1</u>

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