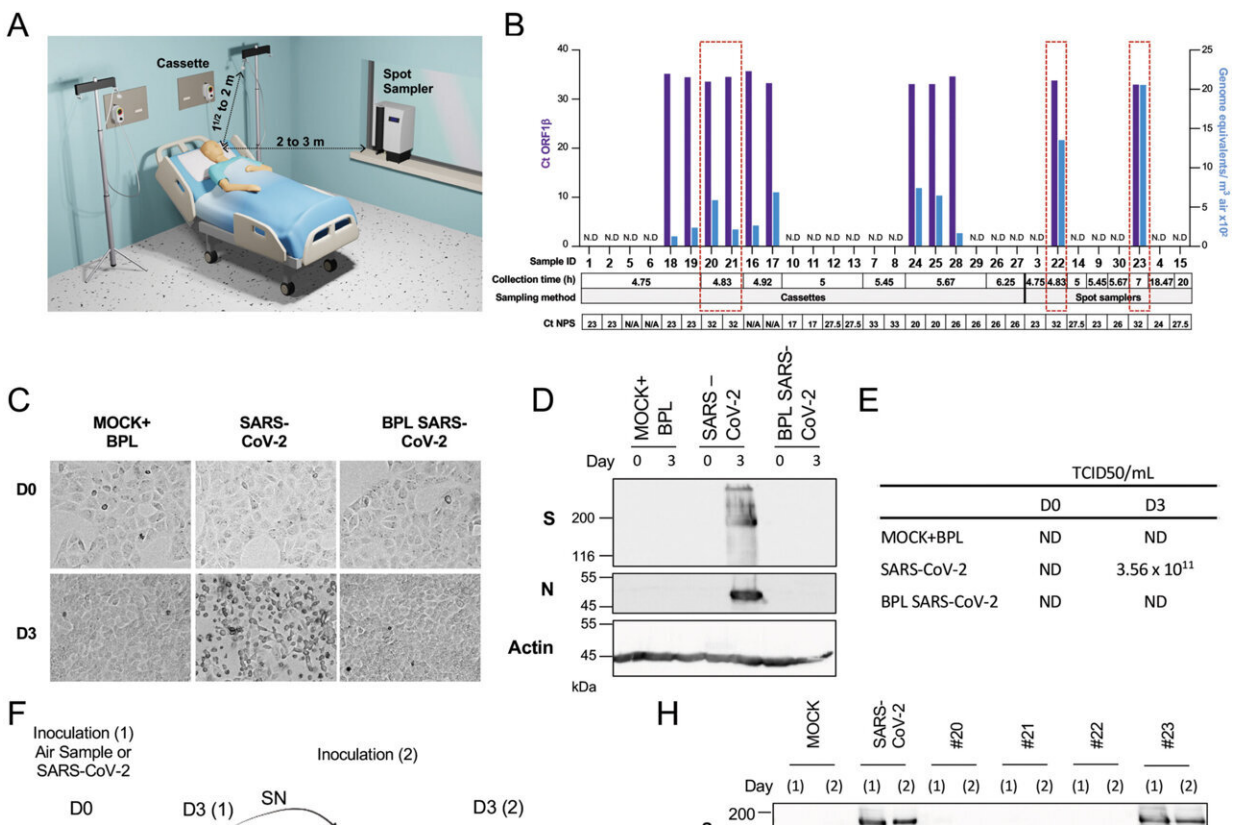


Isolating infectious SARS-CoV-2 from hospital room air samples that were kept frozen for more than a year

April 19 2023, by Bruno Geoffroy



A) Hospital room of patient with COVID-19 layout depicting the position of the sampling devices and the distance to the head of the patient. SKC 37mm closed-face cassettes allow efficient cut off for sampling aerosol and not droplets. They were used pointing downward to prevent accidental contamination by droplets. The Series 110A Liquid Spot Sampler Spot Sampler collects small-size aerosols of 5nm to 10µm. B) Quantification of SARS-CoV-2 ORF1β RNA by RT-qPCR

(Ct, violet bars, left y-axis) and extrapolated genomes equivalent/m³ air (blue bars, right y-axis) in the 30 collected air samples. Red rectangles indicate the four samples that were analyzed for the presence of infectious virus in cell culture. Ct obtained from nasopharyngeal swabs (NPS) for each sample are indicated. C-E) Test experiment to evaluate the cytopathic effects (CPE), expression of Spike (S) and Nucleocapsid (N) and de novo virions production occurring in VERO E6 cells infected with 150 pfu of replicating and β propiolactone (BPL)-inactivated SARS CoV-2. Parameters were evaluated at day 0 (D0) and day 3 (D3) of infection. C) CPE was detected by brightfield microscopy. D) Cellular expression of SARS-CoV-2 S and N proteins was detected by immunoblot using polyclonal anti-SARS-Related Coronavirus 2 Spike Glycoprotein (BEI resources, cat#NR-52947) and SARS Nucleocapsid protein antibody (Novus Biologicals, cat#NB100-56576). E) Production of de novo virions in the supernatant of infected cells was quantified by the median tissue culture infectious dose (TCID₅₀) calculation method. F–I) Analysis of air samples collected in COVID-19 patients hospital rooms. F) Experimental scheme used to culture SARS-CoV-2 from air samples. Air samples were used as inoculum on VERO E6 cells. After 3 days of infection, the supernatant was collected and used to inoculate fresh VERO E6 cells. Control infection was performed with 150 pfu SARS-CoV2/SB2 isolate. CPE (G), cell expression of S and N proteins (H) and production of de novo virions (I) were monitored. ND: not detectable. N.A: not available. Credit: *Clinical Microbiology and Infection* (2023). DOI: 10.1016/j.cmi.2023.03.019

Quebec scientists have succeeded in isolating infectious particles of the SARS-CoV-2 virus from air samples collected from hospital rooms of COVID-19 patients and kept frozen for more than a year, a new study shows.

The research was done by a team led by Nathalie Grandvaux, a researcher at the CHUM Research Centre (CRCHUM) and professor at Université de Montréal, in collaboration with the teams of Caroline Duchaine (Université Laval) and Yves Longtin (McGill University).

Published in *Clinical Microbiology and Infection*, the study provides insight into a scientific field that has been little explored since the beginning of the pandemic.

"In our study, we show through an experimental model that it is possible to isolate and cultivate [infectious viruses](#) from [air samples](#)," said Nathalie Grandvaux. "This is possible even if samples were collected over a year ago then frozen and stored before cell culture."

If public health authorities, including the World Health Organization, were slow to recognize airborne transmission of the [virus](#), it is in part due to limited scientific evidence of the presence of infectious [virus particles](#) in aerosols.

"These studies are difficult to conduct," said Nathalie Grandvaux, "because you need to preserve the infectivity of the virus during collection, use the right cell culture techniques and have access to a level 3 containment laboratory, like the laboratory at the CRCHUM."

A unique method

Audray Fortin, a researcher on Nathalie Grandvaux's team and the first author of the study, developed a unique cell culture method to amplify traces of viruses collected by Caroline Duchaine, a Canadian bioaerosol specialist, and her team.

That was another scientific feat, requiring samplers—a series of collection devices placed inside the rooms of patients with COVID-19—that were used to maintain the virus infectivity and protect it during storage.

In all, 30 samples were collected during the study from 10 different rooms of patients with COVID-19, then stored frozen in a biobank for

14 months.

Using the aerosol samples from one patient's room, the team of scientists was able to identify the presence of replicating virus particles.

"Thanks to our method, it is yet possible to retrospectively evaluate the presence of infectious SARS-CoV-2 in samples collected during the different waves of the pandemic," said Nathalie Grandvaux. "This data will help us better understand the importance of airborne transmission of the virus and implement adapted preventive strategies."

Better prepared for the next pandemic

The findings can be used to prepare for the next pandemic, be it SARS-CoV-2 or another respiratory virus, she added.

"Our research should increase awareness of airborne infectious viruses. It argues for the importance of targeting airborne transmission in personal and collective protection measures, including by improving [indoor air quality](#)."

The collection and culture method can also be adapted to closed environments other than hospitals, such as schools, to test air quality and evaluate the effectiveness of protection measures against airborne transmission of viruses.

More information: Audray Fortin et al, Detection of viable SARS-CoV-2 in retrospective analysis of aerosol samples collected from hospital rooms of patients with COVID-19, *Clinical Microbiology and Infection* (2023). [DOI: 10.1016/j.cmi.2023.03.019](https://doi.org/10.1016/j.cmi.2023.03.019)

Provided by University of Montreal Hospital Research Centre

Citation: Isolating infectious SARS-CoV-2 from hospital room air samples that were kept frozen for more than a year (2023, April 19) retrieved 25 April 2024 from

<https://medicalxpress.com/news/2023-04-isolating-infectious-sars-cov-hospital-room.html>

This document is subject to copyright. Apart from any fair dealing for the purpose of private study or research, no part may be reproduced without the written permission. The content is provided for information purposes only.