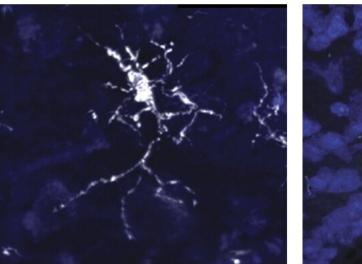
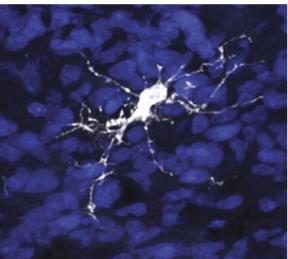


New way to model human brain immune cells

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Left: An immune brain cell (microglia) in a human brain. Right: Microglia in the novel organoid model with a human-brain-like environment. The cells are almost indistinguishable. Credit: Salk Institute

Situated at the intersection of the human immune system and the brain are microglia, specialized brain immune cells that play a crucial role in development and disease. Although the importance of microglia is undisputed, modeling and studying them has remained a difficult task.

Unlike some <u>human cells</u> that can be studied outside the body or in nonhuman models, human microglia are difficult to study when removed



from the human <u>brain</u> environment. To overcome this barrier, Salk scientists developed an organoid model—a three-dimensional collection of cells that mimics features of human tissues.

This model allows researchers to study human microglial development and function for the first time in living human-derived tissue. Further, the scientists examined patient-derived microglia from children with macrocephalic autism spectrum disorder (a condition where infant head circumference is greater than 97% of other infants') to determine whether brain environment influences the development of more reactive microglia.

The findings, published in *Cell* on May 11, 2023, highlight the importance of immune cell and brain interaction and improve the understanding of neurodegenerative and developmental diseases, such as autism spectrum disorder and Alzheimer's disease.

"Outside of the brain environment, microglia lose almost all function and meaning," says Professor Rusty Gage, senior author and holder of the Vi and John Alder Chair for Research on Age-Related Neurodegenerative Disease. "We knew that if we found a way to replicate the human brain environment in an organoid in order to study human microglia, then we would finally have a tool for examining how the heathy and diseased brain influence microglia and, reciprocally, how healthy and diseased microglia influence the brain."

Emerging roughly 10 years ago, organoids have become a prevalent tool to bridge the gap between cell and <u>human studies</u>. Organoids can mimic human development and organ generation better than other laboratory systems, allowing researchers to study how drugs or diseases affect human cells in a more realistic setting. Brain organoids are typically grown in culture dishes, but the organoids are structurally and functionally limited by the lack of blood vessels, short survival time, and



inability to sustain diverse cell types (like microglia).

"To create a brain organoid model that contains mature microglia and enables us to research them, we used a novel transplantation technique to create a human-brain-like environment" says co-first author Abed Mansour, a former postdoctoral researcher in Gage's lab and now an assistant professor at the Hebrew University of Jerusalem. "So we could finally make a human brain organoid that had all the features necessary to orchestrate human microglia growth, behavior, and function."

Unlike previous models, the researchers created a human brain organoid that had microglia *and* a human-brain-like environment, which finally allowed them to look at <u>environmental influences</u> on microglia throughout brain development. They found that a characteristic protein called SALL1 appeared as early as eleven weeks into development and served to confirm microglial identity and promote mature function. Additionally, they found that brain environment–specific factors, like the proteins TMEM119 and P2RY12, were necessary for microglia to function.

"Creating a human brain model that can effectively replicate the human brain environment is very exciting," says Associate Professor Axel Nimmerjahn, another author of the study. "With this model, we can finally investigate how human microglia function within the human brain environment."

As the team learned more about microglia, the importance of the relationship between brain environment and microglia became clear—especially in disease scenarios. The lab <u>previously examined</u> neurons derived from people with autism spectrum disorder and found their neurons grew faster and had more complex branches than neurotypical counterparts. With the new <u>organoid</u> model, the team could ask whether those neuronal differences altered the brain environment



and influenced microglia development.

To do so, they compared microglia derived from skin samples from three individuals with macrocephalic autism spectrum disorder versus three neurotypical individuals with macrocephaly. The researchers found that individuals with autism spectrum disorder exhibited the neuronal differences the team had previously noted, and that the microglia were influenced by those differences in their growth environment. Because of this neuron-dependent environmental change, the microglia became more reactive to damage or intruders—a finding that may explain the brain inflammation observed in some individuals with <u>autism spectrum disorder</u>.

Since this was a preliminary study with a <u>small sample size</u>, the team plans to examine more microglia from additional people in the future to verify their findings. They also aim to expand their research to study other developmental and neurodegenerative diseases to see how <u>microglia</u> are contributing to disease onset.

"Rather than deconstruct the brain, we decided to construct it ourselves," says co-first author Simon Schafer, a former postdoctoral researcher in Gage's lab and now an assistant professor at Technical University of Munich. "By building our own brain model we can work from the bottom up and see solutions that may be impossible to see from the top down. We are eager to continue improving on our model and unraveling the relationship between the brain and immune system."

More information: Fred H. Gage, An in vivo neuroimmune organoid model to study human microglia phenotypes, *Cell* (2023). <u>DOI:</u> 10.1016/j.cell.2023.04.022.

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