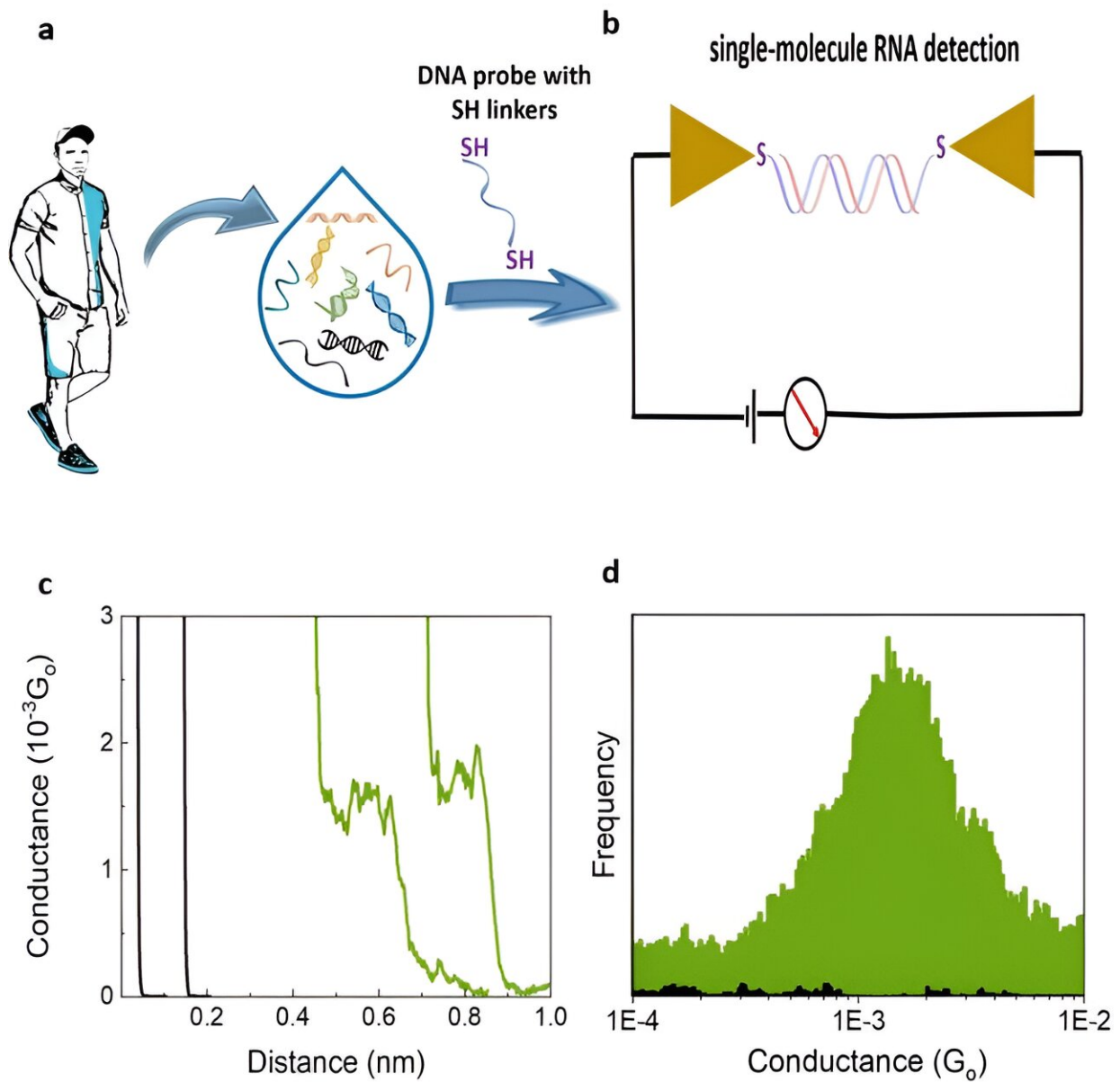


Electrical detection of RNA cancer biomarkers at the single-molecule level

August 17 2023, by Thamarasee Jeewandara



Detection of cancer biomarkers using the STMBJ single-molecule conductance approach. (a) Liquid biopsy samples contain circulating nucleic acids that can be detected with a complementary DNA probe capable of binding to STM electrodes. (b) STMBJ detection of the hybridized biomarker, resulting in a step in the conductance–distance signal. (c) Example of collected conductance–distance curves. Control phosphate buffer solution (black), green: example raw data curves collected from the G12C sample. (d) Histogram built from curves with steps (black: Phosphate buffer blank, green: G12C RNA sample), 118 from a total 5399 curves acquired in the experiment. Credit: *Scientific Reports*, doi: <https://doi.org/10.1038/s41598-023-39450-6>

Early screening methods offer a promising avenue to reduce mortality rates of cancer, notably through the analysis of biomarkers in liquid biopsies. Biologists have shown how the electrical detection of nucleic acids at the single molecule level can facilitate such applications.

In a new paper published in *Scientific Reports*, Keshani G. Gunasinghe and a team of scientists in chemistry at the University of Massachusetts, U.S., studied the electrical detection of RNA [cancer](#) biomarkers for cancer screening applications as a proof-of-concept electrical biosensor.

Sequence-sensitive electrical conductance of the instrument allowed the discrimination of mutants from the wild-type. Their specificity showed the sensitivity of biosensors down to an individual molecule with a high signal-to-noise ratio. The outcomes pave the way to engineer miniaturized single-molecule electrical biosensors that are groundbreaking for cancer screening.

Cancer screening with scanning tunneling microscopy-assisted break junctions (STMBJ)

The World Health Organization lists cancer as [a leading cause of mortality](#); these statistics can be evaded through the [early detection of the disease](#) via non-invasive analysis of liquid biopsies. Liquid biopsies target cancer-specific biomarkers in blood or saliva including [tumor cells](#) or [circulating tumor nucleic acids](#) such as circulating tumor DNA and RNA variants—ctDNA and ctRNA.

It is challenging to detect ct-nucleic acids (ctNA) in liquid biopsies due to their low concentrations and the low frequency of mutations when compared to the wild-type. Bioengineers have sought [advances in nanoscience and nanotech](#) to help address these challenges, which include [scanning tunneling microscopy-assisted break junctions](#) (STMBJ) that life scientists had recently used to detect the first single-molecule, by [identifying RNA from E.coli](#) strains.

This approach can be extended to detect cancer [biomarker](#) RNA sequences from human liquid biopsies. The method can detect small amounts of RNA biomarkers in solution via electrical means to obtain sequence-specific fingerprints that are sensitive to single-point mutations.

In this work, Gunasinghe et al. developed a proof-of-concept STMBJ method as a biosensor. The outcomes highlighted the first single molecule conductance results for sequences of human origin.

a

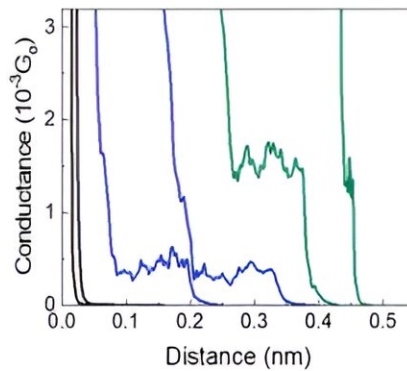
G12C mismatch 18nt

SH-5' G12C 18nt DNA probe 3'-SH
 GGAGCTTGTGGCGTAGGC
 CCUCGACCA~~C~~CCGAUCCG
 3' Wild type 18nt RNA target 5'

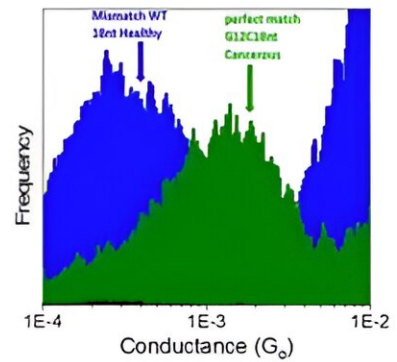
G12C perfect match 18nt

SH-5' G12C 18nt DNA probe 3'-SH
 GGAGCTTGTGGCGTAGGC
 CCUCGACCA~~C~~CCGAUCCG
 3' G12C 18nt RNA target 5'

b



c



d

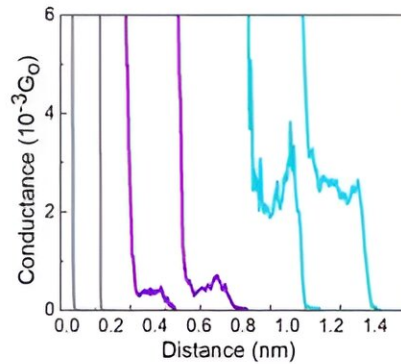
G12V mismatch 18nt

SH-5' G12V 18nt DNA probe 3'-SH
 GGAGCTGTTGGCGTAGGC
 CCUCGACCA~~C~~CCGAUCCG
 3' Wild type 18nt RNA target 5'

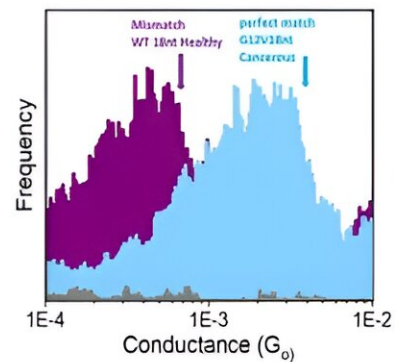
G12V perfect match 18nt

SH-5' G12V 18nt DNA probe 3'-SH
 GGAGCTGTTGGCGTAGGC
 CCUCGACCA~~C~~CCGAUCCG
 3' G12V 18nt RNA target 5'

e



f

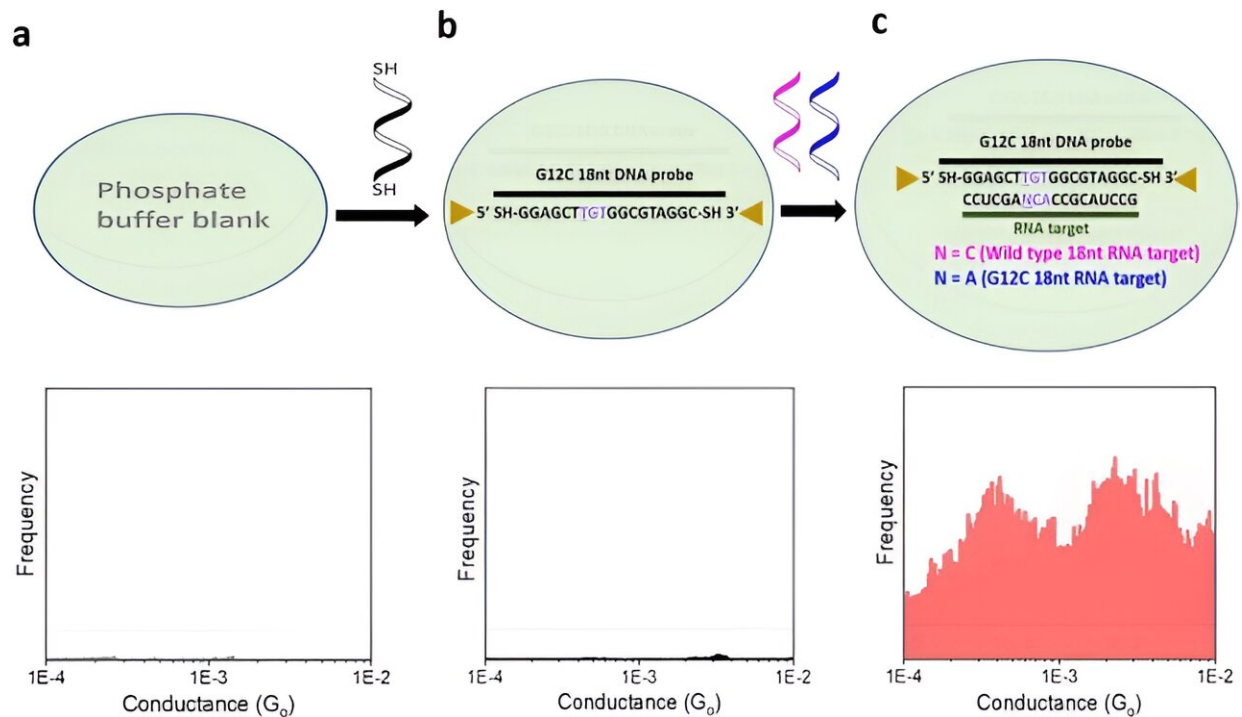


Electrical detection of KRAS mutations. (a) Sequences for G12C 18nt mismatch and perfect match. (b) Example conductance versus distance curves (Black: blank, Blue: mismatch, Green: perfect match). (c) Histograms for G12C mismatch (379 from a total 5203 curves acquired) and perfect match (212 from a total 13,958 curves acquired) overlapped with phosphate buffer blank (Blue: G12C 18nt mismatch, Green: G12C 18nt perfect match, Black: phosphate buffer blank). (d) Sequences for G12V 18nt mismatch (healthy) and perfect match (cancerous). (e) Raw conductance versus distance curves (Black: blank, Purple: mismatch, Light blue Blue: perfect match). (f) Histograms for G12V mismatch (75 from a total 5024 curves acquired) and perfect match (116 from a total 10,159 curves acquired) overlapped with phosphate buffer blank (Purple: G12V 18nt mismatch, Light blue: G12V 18nt perfect match, Gray: phosphate buffer blank). Credit: *Scientific Reports*, doi: <https://doi.org/10.1038/s41598-023-39450-6>

Mechanism of action

The team detected a liquid biopsy sample containing multiple ctDNA by targeting it with specific dithiol-modified DNA probes. They then electrically detected the single molecule RNA cancer biomarkers by hybridizing the DNA probe with the target RNA to [create a DNA: RNA hybrid](#) that bound to the scanning tunneling microscopy electrodes. The outcomes formed closed circuit biomolecular electronics to measure electrical fingerprints in the form of STMBJ [conductance-distance curves](#) to represent steps of molecular conductance.

Gunasinghe and colleagues performed STMBJ experiments to electrically detect RNA cancer biomarkers. They measured the electrical conductance of two mutant variants and differentiated the mutation from the wild-type sequence simultaneously in solution. Using titration experiments, they showed a limit of detection with proof-of-concept electrical biosensors at the single molecule level with a high signal-to-noise ratio.



High specificity of the STMBJ approach for cancer detection: In situ hybridization for perfect match G12C. (a) Histogram for blank phosphate buffer (3 from a total 1150 curves acquired). (b) Histogram for G12C DNA probe (3 from a total 661 curves acquired). (c) Conductance histogram including both G12C and wild-type DNA:RNA hybrids (111 from a total 5631 curves acquired). Credit: *Scientific Reports*, doi: <https://doi.org/10.1038/s41598-023-39450-6>

Designing biomolecular electronic DNA probes through bioinformatics

Genomics studies on cancer biomarkers include the [Pancancer Analysis of Whole Genomes](#), the Consortium of the International Cancer Genome Consortium, and the [Cancer Genome Atlas](#) that has assessed whole cancer genomes [from several tumor types](#).

This has created an opportunity to identify potential mutant sequences for RNA cancer biomarkers to be observed in [liquid biopsy samples](#). The Kirsten rat sarcoma is a well-known oncogene with a high mutation rate in several cancers frequently found in pancreatic ductal adenocarcinoma, [colorectal cancer and skin melanoma](#).

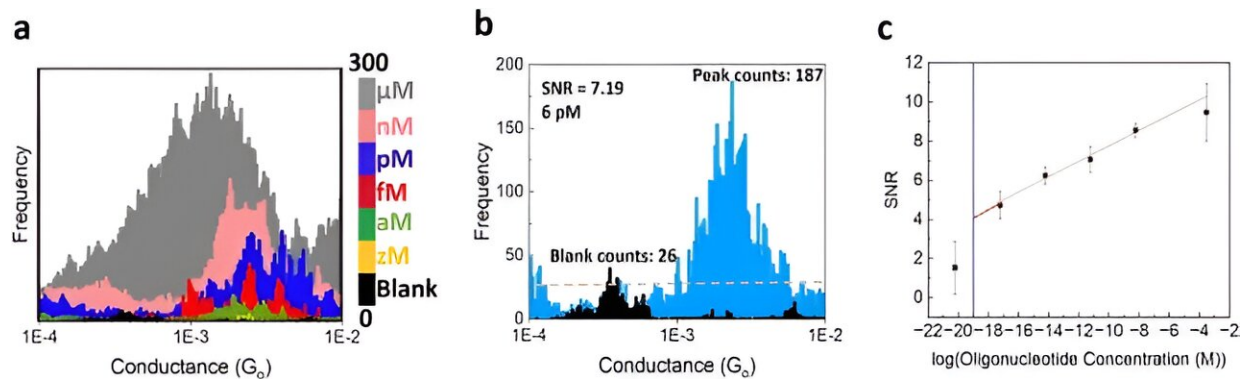
Such mutations typically contain a single base-level mutation occurring at codon 12 present in 46% of all lung adenocarcinomas known as G12, while this is present in a variety of [colorectal and pancreatic cancers](#). The team focused on two specific mutations that covered a variety of cancer types with [promising future screening approaches](#).

Electrical detection of single-molecule cancer biomarkers

Using the scanning tunneling microscopy break-junction method (STMBJ), Gunasinghe et al. measured the charge transport conductance values of individual mutant RNA molecules.

They recorded a range of conductance-distance curves. In the experimental setup, they modified the DNA probes with [thiol binding groups](#) to facilitate them linking to electrodes to hybridize with a complementary target RNA sequence and form a conductance step by closing the biomolecular electronics circuit. The sensitivity of the instrument prevented the observation of steps when there were no molecules bound between the two molecules.

This biosensing strategy allowed them to differentiate the mutants from the wild-type variants. The outcomes generated the first single-molecule measurements of an RNA sequence of human origin, while further elucidating it to be the first electrical detection of individual cancer biomarker molecules.



High sensitivity of the STMBJ approach for cancer detection. (a) Conductance histograms for G12C titration experiments (concentration varies from 300 μM to 0. The control experiment in phosphate buffer solution (black) shows no peaks in the histogram. (b) Limit of Detection (LoD), Example of SNR calculation for a 6 pM concentration sample. (c) Average SNR for each concentration (with a linear fit) to get the concentration for a SNR=3, Blue vertical line: theoretical concentration where a single molecule is present in the sample volume: around 0.1 aM). Credit: *Scientific Reports*, doi: <https://doi.org/10.1038/s41598-023-39450-6>

Specificity and sensitivity of the instrument

In this instance, Gunasinghe and colleagues defined specificity as the ability to discriminate cancer biomarkers from the wild-type sequence. The single-molecule electrical biosensor showed high specificity to detect the sequence of interest, allowing future applications to detect multiple disease-related sequences [in parallel to diagnose a variety of diseases](#).

The team assessed the sensitivity of the approach through titration experiments to determine the limits of detection (LOD) of the system and defined the parameters of minimum target concentration. These

outcomes provided a signal-to-noise ratio corresponding to the level of an individual molecule; a new target limit of detection. The titration experiments confirmed the superior sensitivity, and single-molecule nature of the biosensor to pave the way for cancer screening in a miniaturized, automated instrument in the future.

Outlook

In this way, Keshani G. Gunasinghe and colleagues presented a fully electrical proof-of-concept biosensor to detect RNA cancer biomarkers in solution with sensitivity at the level of a single molecule. The study represents a first application of single molecule measurements down to a sequence, for samples originating from humans.

Using the instrument, they efficiently identified the cancer biomarkers from the wild-type counterparts with a high signal-to-noise ratio. The extremely specific system simultaneously detected and differentiated multiple human RNA targets as healthy wild-type and mutant variants at a high specificity and sensitivity. The nanobiosensor has a promising future as an inexpensive, highly sensitive and label-free miniaturized device for early-stage cancer screening of liquid biopsies with future applications in biomedicine.

More information: Keshani G. Gunasinghe Pattiya Arachchillage et al, Electrical detection of RNA cancer biomarkers at the single-molecule level, *Scientific Reports* (2023). [DOI: 10.1038/s41598-023-39450-6](https://doi.org/10.1038/s41598-023-39450-6)

Yuanhui Li et al, Detection and identification of genetic material via single-molecule conductance, *Nature Nanotechnology* (2018). [DOI: 10.1038/s41565-018-0285-x](https://doi.org/10.1038/s41565-018-0285-x)

Citation: Electrical detection of RNA cancer biomarkers at the single-molecule level (2023, August 17) retrieved 28 April 2024 from <https://medicalxpress.com/news/2023-08-electrical-rna-cancer-biomarkers-single-molecule.html>

This document is subject to copyright. Apart from any fair dealing for the purpose of private study or research, no part may be reproduced without the written permission. The content is provided for information purposes only.