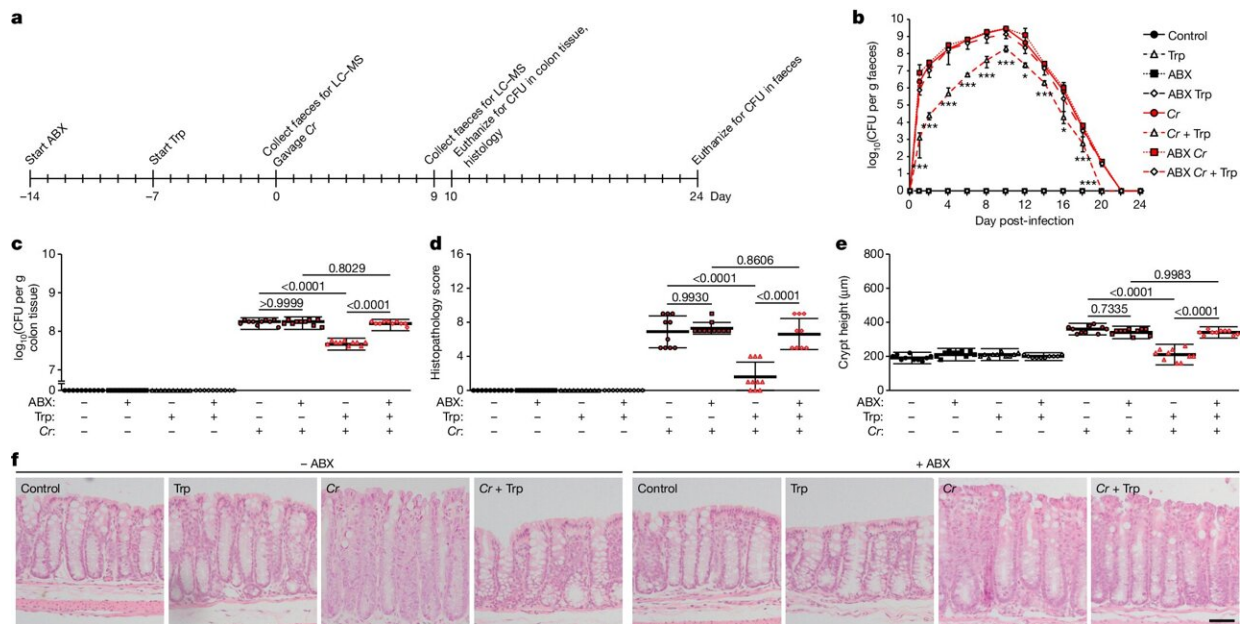


Tryptophan in diet and gut bacteria protect against *E. coli* infection, study shows

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Dietary Trp protects against infection with *C. rodentium* strain DBS100 in a mouse model of EHEC infection. Credit: *Nature* (2024). DOI: 10.1038/s41586-024-07179-5

Gut bacteria and a diet rich in the amino acid tryptophan can play a protective role against pathogenic *E. coli*, which can cause severe

stomach upset, cramps, fever, intestinal bleeding and renal failure, according to a study [published](#) March 13 in *Nature*.

The research reveals how dietary tryptophan—an amino acid found mostly in [animal products](#), nuts, seeds, whole grains and legumes—can be broken down by gut bacteria into small molecules called metabolites. It turns out a few of these metabolites can bind to a receptor on gut epithelial (surface) cells, triggering a pathway that ultimately reduces the production of proteins that E. coli use to attach to the gut lining where they cause infection. When E. coli fail to attach and colonize the gut, the pathogen benignly moves through and passes out of the body.

The research describes a previously unknown role in the gut for a receptor, DRD2. DRD2 has otherwise been known as a dopamine (neurotransmitter) receptor in the central and peripheral nervous systems.

"It's actually two completely different areas that this receptor could play a role in, which was not appreciated prior to our findings," said Pamela Chang, associate professor of immunology in the College of Veterinary Medicine and of [chemical biology](#) in the College of Arts and Sciences. "We essentially think that DRD2 is moonlighting in the gut as a microbial [metabolite](#) sensor, and then its downstream effect is to help protect against infection."

Samantha Scott, a postdoctoral researcher in Chang's lab, is first author of the study, titled "Dopamine Receptor D2 Confers Colonization Resistance via Microbial Metabolites."

Now that Chang, Scott and colleagues have identified a specific pathway to help prevent E. coli infection, they may now begin studying the DRD2

receptor and components of its downstream pathway for therapeutic targets.

In the study, the researchers used mice infected with *Citrobacter rodentium*, a bacterium that closely resembles *E. coli*, since certain pathogenic *E. coli* don't infect mice. Through experiments, the researchers identified that there was less pathogen and inflammation (a sign of an active immune system and infection) after mice were fed a tryptophan-supplemented diet.

Then, to show that gut bacteria were having an effect, they gave the mice antibiotics to deplete microbes in the gut, and found that the mice were infected by *C. rodentium* in spite of eating a tryptophan diet, confirming that protection from tryptophan was dependent on the gut bacteria.

Then, using [mass spectrometry](#), they ran a screen to find the chemical identities of tryptophan metabolites in a gut sample, and identified three such metabolites that were significantly increased when given a tryptophan diet. Again, based on pathogen levels and inflammation, when these three metabolites alone were fed to the mice, they had the same protective effect as giving the mice a full tryptophan diet.

Switching gears, the researchers used bioinformatics to find which proteins (and receptors) might bind to the tryptophan metabolites, and from a long list they identified three related receptors within the same family of dopamine receptors. Using a human intestinal cell line in the lab, they were able to isolate receptor DRD2 as the one that had the protective effect against infection in the presence of [tryptophan](#) metabolites.

Having identified the metabolites and the receptor, they analyzed the downstream pathway of DRD2 in human gut [epithelial cells](#). Ultimately, they found that when the DRD2 pathway was activated, the host's ability

to produce an actin regulatory protein was compromised. *C. rodentium* (and *E. coli*) require actin to attach themselves to gut epithelial cells, where they colonize and inject virulence factors and toxins into the cells that cause symptoms. But without actin polymerization they can't attach and the pathogen passes through and clears.

The experiments revealed a new role of dopamine receptor DRD2 in the gut that controls actin proteins and affects a previously unknown pathway for preventing a pathogenic bacteria's ability to colonize the gut.

Jingjing Fu, a former postdoctoral researcher in Chang's lab, is a co-author.

More information: Samantha A. Scott et al, Dopamine receptor D2 confers colonization resistance via microbial metabolites, *Nature* (2024). DOI: [10.1038/s41586-024-07179-5](https://doi.org/10.1038/s41586-024-07179-5)

Provided by Cornell University

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