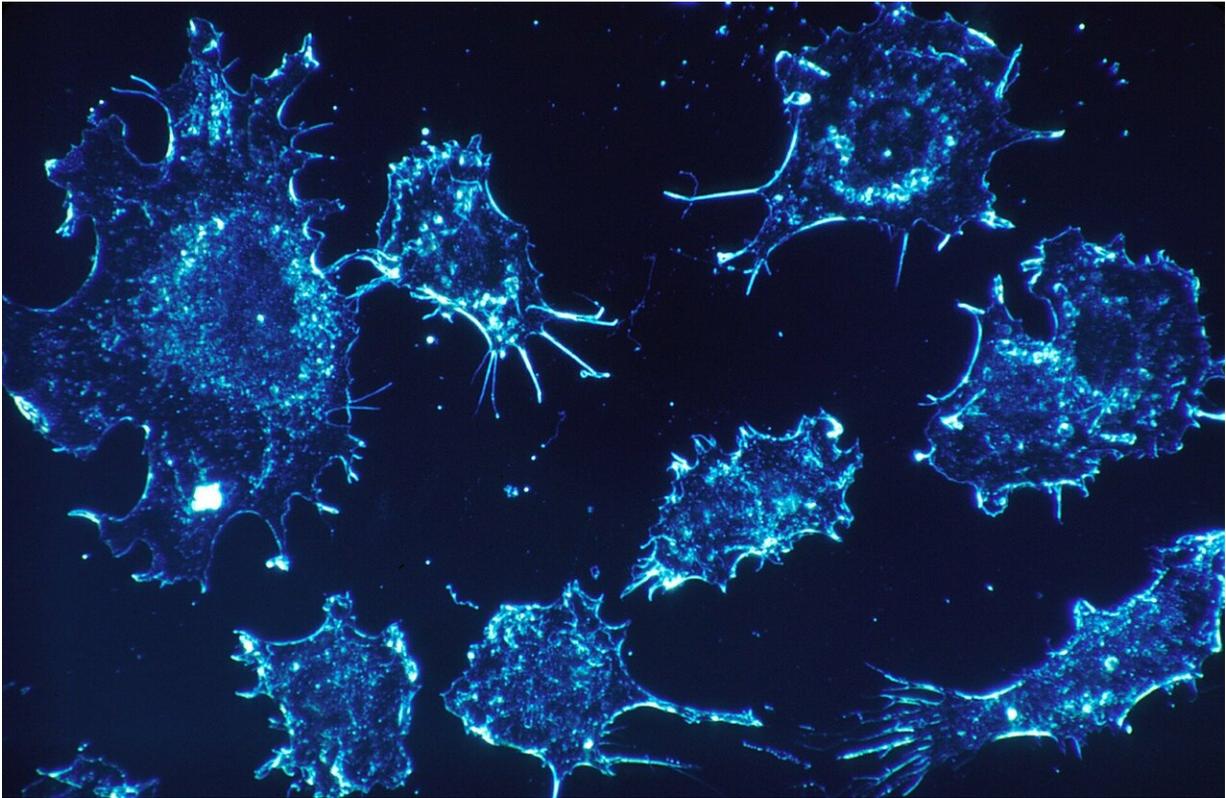


# Are all cancer cells the same?

February 21 2019, by Rodolphe Marie, Jonas N. Pedersen, Henrik Flyvbjerg, Anders Kristensen

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Credit: CC0 Public Domain

Take two cancer cells and compare their genomes. Surprisingly, they can be quite different. This genetic variation is one of the hallmarks of cancer, and one reason why treating cancer is so hard.

If a tumour is made up of [cells](#) with many different genomes, a single drug might not kill them all. But knowing the [genetic variation](#) can help clinicians to develop targeted treatments.

As part of our research, we have trapped a single cancer cell on a plastic device (see figure), extracted its DNA, and produced a coarse-grained picture of its sequence. Our results are published in the journal *Proceedings of the National Academy of Sciences (PNAS)*.

## Optical mapping of DNA from a single cell

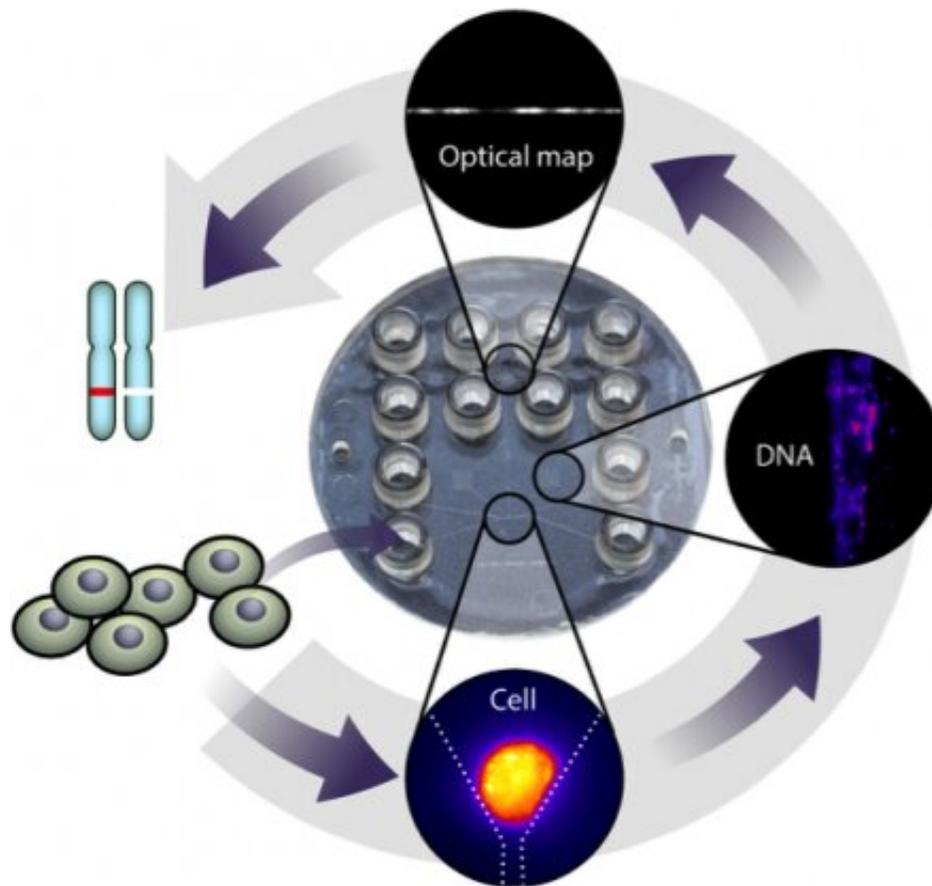
We used a technique called [optical mapping](#), which provides large-scale information about the [genome](#). It works like a map of the world with forests, lakes, and mountains, but without the finer details like roads, houses and small cities.

By comparing optical maps from a [single cell](#) with a reference map for the average human cell, we can pinpoint differences between them. This information can reveal genomic heterogeneity within a tumour and may even indicate how cells have developed into tumours.

Optical mapping of DNA from a single cell consists of four steps:

- First we capture a cell and extract long fragments of its DNA.
- Then we dye the DNA fragments with a [fluorescent dye](#).
- Then we heat the DNA [molecules](#). The dye sticks better in some places than others, depending on the DNA sequence. This leaves a barcode-like pattern on the molecules.
- Finally, we stretch and image the molecules under a microscope to read the 'barcodes'.

We developed a low-cost plastic device that integrates all four steps. The steps are shown in the figure below.



Human cells are loaded onto a single-use plastic device. A single cell is captured, and its DNA is extracted and stained with a fluorescent dye. The DNA is then heated to create a 'barcode', i.e. a pattern of fluorescence that depends on its specific DNA sequence. Individual DNA fragments are stretched, and their barcodes are imaged and analysed. Comparison with a reference genome shows genetic changes in the specific cell that the DNA came from.

The 'barcode' works like a fingerprint: It identifies which part of the reference genome a DNA molecule is closely related to. And it can even reveal differences between the imaged DNA molecule and the reference genome.

## Sequencing versus optical mapping

So why use optical mapping and not just the usual DNA sequencing? Conventional DNA sequencing methods have the advantage that they have single-base-pair resolution, meaning that every base pair in a DNA molecule can be identified. Given enough material, researchers can sequence the entire genome from a human cell – approximately 6 billion bases – in a few days. But it is challenging to sequence a single copy of a human genome. And that is all we have when we start with a single cell.

One of the challenges is that the usual DNA sequencing methods require multiple copies of the genome. Since there is only one copy of the genome in each cell, the first step is to copy the genome multiple times. This is called DNA amplification and is routine chemistry. But copying errors occur occasionally, which obscure the results.

Another challenge is that each copy of the DNA molecule is randomly cut into smaller pieces, just a few hundred base pairs long. These pieces are then sequenced. The results, the so-called 'reads', are then assembled into a full genome by matching partially overlapping reads.

It is also very difficult to detect structural variations, such as repeating patterns or inserted/missing genomic elements that are longer than the few hundred-base-pair read lengths. But it is just this type of structural information that may prove useful for the selection of cancer therapies.

There is, at least in theory, a more obvious and efficient way to read the DNA sequence. The genome is coded on 48 DNA molecules in two-nanometres-thick fibres up to eight centimetres long. So, why not just read the DNA sequence from one end to the other?

Optical mapping almost does this: It provides a rough 'fingerprint' of the underlying DNA sequence of DNA molecules up to 1 million base pairs

long. That is much longer than the short read lengths of DNA sequencing. And optical mapping also avoids the amplification steps needed for DNA sequencing.

The long fragments make it possible to detect structural variations that are from a few kilo-base pairs up to several hundred kilo-base pairs. Smaller variations can be detected with DNA sequencing.

So the two methods (sequencing and mapping) are complementary. And the same DNA molecule can, in principle, be both optically mapped and then sequenced.

## **Toward a more efficient and personalised cancer treatment**

The ability to sequence the genome at the single-cell level could lead to more efficient and individualised cancer treatments. But as we explained above sequencing the DNA from a single-cell is still demanding using current sequencing techniques.

We have demonstrated for the first time that optical mapping can detect large-scale genetic variations in a DNA molecule taken from a single cell. And we do it without the amplification steps required by DNA sequencing methods.

The sample preparation was done entirely on a single-use lab-on-a-chip device, starting from a single cell and finishing with useful genomics data. This is an important technological aspect of our work, as the device reduces the use of expensive chemicals and minimises the risk of contamination.

To be clear, our work is still in the research phase. Our device is not yet

ready for use in hospitals, and the challenge now is to increase the throughput, so we can analyse more DNA molecules at a time. The goal is to map all the DNA from a single cell.

**More information:** Rodolphe Marie et al. Single-molecule DNA-mapping and whole-genome sequencing of individual cells, *Proceedings of the National Academy of Sciences* (2018). [DOI: 10.1073/pnas.1804194115](https://doi.org/10.1073/pnas.1804194115)

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